

Potential Impacts to Biodiversity from Proposed Lithium Extraction in Nevada and California

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The Nature
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COVER Lithium mining in Nevada © Jassen Todorov/TNC Photo Contest 2022; THIS PAGE The Virgin River © Chip Carroon/TNC; Pronghorn © Chip Carroon/TNC; Desert tortoise © Dana Wilson/BLM; Jackson Basin in Esmeralda County, Nevada has several potential lithium projects © Michael Clifford/TNC

Introduction

Climate change and biodiversity loss are two of the greatest threats to ecosystems and human health (Chapin III et al. 2000; Diaz et al. 2019). The impacts of climate change can drive species range shifts (Chen et al. 2011), cause local extirpations (Sinervo et al. 2010), and increase species extinctions (Pounds et al. 2006). At the same time, the loss of key species can disrupt vital ecosystem services, such as pollination, nutrient cycling, and carbon sequestration, creating a feedback loop that intensifies both climate change and biodiversity loss (Chapin III et al. 2000; Cardinale et al. 2012; Diaz et al. 2019). Thus, climate change and biodiversity loss are strongly connected and solutions to maintain biodiversity such as restoring habitat, or maintaining landscape connectivity can mitigate climate change through carbon uptake and storage (Pörtner et al. 2023; von Jeetze et al. 2023).

Addressing these interconnected challenges necessitates comprehensive strategies that combine emissions reduction, conservation efforts, habitat restoration, and sustainable land-use practices to protect habitats and species. However, climate change solutions may come at the expense of biodiversity if appropriate planning is not done to avoid developing where the most sensitive species and habitats occur (Sonter et al. 2020; Sonter et al. 2023; Wu et al. 2023). Solutions focused only on mitigating climate change and reducing greenhouse gases may undermine biodiversity by not taking a holistic perspective of the two issues. Present rates of extinction are estimated at 1000-times higher than background-levels (Primm et al. 2014), which is largely driven by habitat loss (Ceballos et al. 2015; Diaz et al. 2019; Powers and Jetz 2019). Rates of extinction vary by geography and taxa, but the highest rates occur in small range and small population species (Primm et al. 2014; Staude et al. 2020). Extinction risk is further exacerbated by climate change (Sinervo et al. 2010).

Actions must be taken to limit the amount of greenhouse gas emissions to keep global temperatures below the often agreed upon warming goal of 1.5° C (IPCC 2018). To keep global average temperatures from crossing critical targets, decreasing the use of fossil fuels is necessary. Shifting from fossil fuel-based energy and internal combustion engines to renewable energy and electric vehicles (EVs) will reduce carbon emissions. However, the energy shift will significantly increase the need to mine additional minerals in order to meet material demands for renewable energy generation (e.g., photovoltaic cells and wind turbines) (Sonter et al. 2020), transmission, and storage beyond what is currently mined and extracted (Jowitt and McNulty 2021). In particular, lithium is an important mineral in the production of batteries, specifically lithium-ion batteries, which are relatively light in weight, making them well suited for EVs, mobile phones, and other domestic uses. Lithium-ion batteries are also highly energetic and hold long charges relative to other metals; 80% of global lithium production in 2022 was used to manufacture batteries (USGS 2023). To replace the global supply of internal combustion engines with EVs, it has been estimated that the amount of lithium produced will need to increase approximately 40-times (Herrington 2021; Haddad et al. 2023).

Three global regions currently dominate lithium production, including Australia (47% of global lithium production by weight), South America (35%), and China (15%) (Bradley et al. 2017; Parker et al. 2022). At present the US has one operational lithium extraction facility that is located in Nevada (Bradley et al. 2017). Large concentrations of lithium deposits have been identified throughout the contiguous US (Bradley et al. 2017), but Nevada and California contain approximately 89% of known reserves (Parker et al. 2022). Lithium deposits in the US are found in several forms, including brines, clays, and granitic rocks.

The different methods used to extract lithium from each source material vary in their potential environmental impacts (Vera et al. 2023; Parker et al. 2024). Proposed methods of lithium extraction in California and Nevada include brine extraction, open pit mining, and clay surface mining (Parker et al. 2022).

Underground brine reservoirs (salars or playas) are the focus of 75% of the proposed lithium projects in the region (Parker et al. 2022). In the evaporative concentration method, brine is pumped to the surface where it is concentrated in a series of evaporation ponds (Vera et al. 2023). Evaporation ponds typically require warm, dry climates, and result in a large surface disturbance of hundreds to thousands of hectares. Additionally, the evaporative process consumes large amounts of water (Vera et al. 2023), and produces large volumes of waste (Flexer et al. 2018). Direct Lithium Extraction (DLE) is an emerging technology that has yet to be implemented at industrial scales, but is a process where brines are pumped to the surface, and lithium is extracted in a closed system through mechanical, chemical, electrical, or other methods and the lithium-depleted brines are reinjected back into the brine deposits (see Vera et al. 2023 for detailed review). DLE likely has a small land disturbance area relative to other mining methods (Parker et al. 2022), however the freshwater requirements may be significant (Vera et al. 2023). Lithium extracted from hard rock deposits rely on open pit mining, usually of pegmatites (spodumene). Conceptually, the surface disturbances of open pit mining for lithium are similar to other open pit mines where the ore is extracted, crushed, and treated to obtain the desired mineral. However, the chemicals used to extract lithium from the ore may be different than those used for other minerals. Groundwater may also be intercepted during mine development and would require removal to access the lithium.

Lithium mine claims in California and Nevada are located in the desert regions where there is low annual precipitation. These desert ecosystems are mostly intact landscapes with pockets of high biodiversity and unique species adapted to the extreme climatic conditions (Randall et al. 2010). The region also has many isolated wetlands and groundwater dependent ecosystems (GDEs), which are known to hold outsized importance for biodiversity as many contain rare or endemic species that have been isolated for millennia (Davis et al. 2013). GDEs and the unique species assemblages they support are highly vulnerable to change, and therefore impacts to these systems can serve as

a proxy for environmental impact and biodiversity loss. Furthermore, water scarcity and increasing aridity is a major challenge with many basins that are already over pumped (Parker et al. 2021; Saito et al. 2022). Understanding the impacts to groundwater resources from lithium extraction is important for predicting ecosystem responses to changes in hydrology.

To evaluate the risk of hydrological impacts from lithium mining at a project level, The Nature Conservancy (TNC) contracted the Desert Research Institute (DRI) to develop a framework and checklist (Saftner et al. 2023). The checklist and framework can be used to identify areas of uncertainty with respect to potential impacts to the hydrology, surface water or groundwater on individual projects during the life-cycle of a mine from development through post-closure. The framework and checklist were designed to be used with all available hydrological data (e.g., environmental documents, project plans, reports, etc.) for a project

area. Questions in the framework can lead the user to determine if “red flags” exist in a developer’s plans, or if additional studies or information should be requested.

Here we investigate possible impacts to biodiversity from the development of potential lithium projects in California and Nevada. Parker et al. (2024) provided a broad analysis of potential environmental impacts of this activity at 72 proposed extraction sites in the US. Our analyses use data from Parker et al. (2024) to further focus and analyze the possible impacts to biodiversity in California and Nevada. We specifically focus on the potential impacts to imperiled and vulnerable species (G1-G3 and S1-S3 rank) from potential lithium extraction at two different scales: proposed lithium projects and a broader lithium focus area.



Sarcobatus Flat in Nye County, Nevada has several potential lithium projects © Michael Clifford/TNC

Methods

To evaluate the potential impacts of lithium extraction on biodiversity, we used a geospatial overlay analysis at a fine-scale individual project level, and a broad-scale regional analysis to identify conflict between species occurrence and potential mining operations in California and Nevada. For our fine-scale analysis, we defined “proposed lithium projects”, which were individual mine projects consisting of mine claims based on the “potential lithium projects” identified by Parker et al. (2024).

These included contiguous mine claims operated by a single entity (e.g., company, individual, etc.) that could be grouped and had company plans or filings with the state and/or federal government, and were actively being explored, permitted, and developed. Lithium mine claims lacking a detailed company association were not included in our analyses. Each proposed lithium project area included a 3.2 km (2 mile) buffer around the original claim or claims because it is likely that a developed mine site will have im-

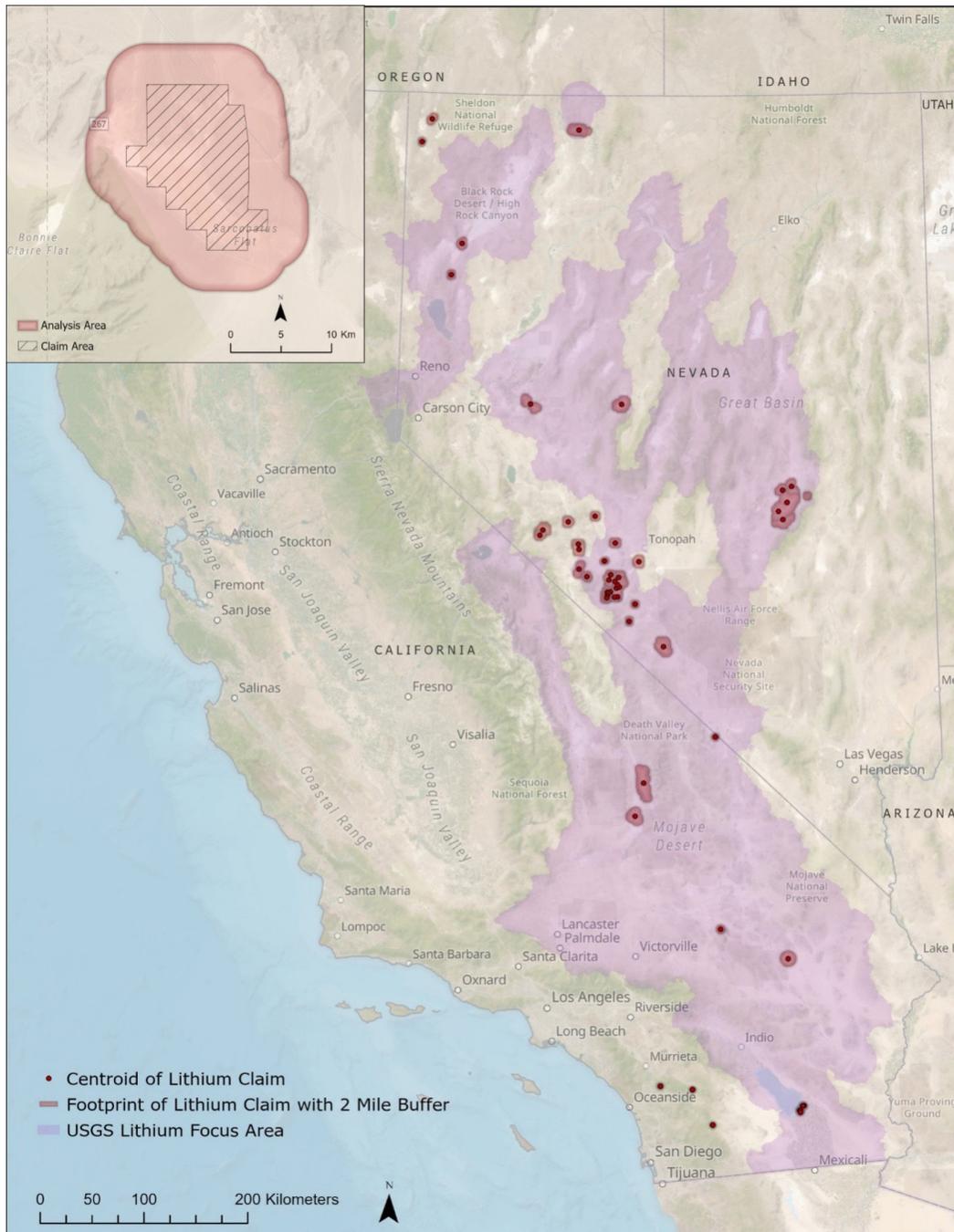


Figure 1. The study area showing individual proposed lithium projects and the lithium focus area. Inset map shows a proposed project, including the claim and buffer area.

pacts beyond the claim boundary in the form of light pollution, dust pollution, access roads, transmission lines, etc.

At the broader-scale we analyzed the potential impacts on biodiversity within the “lithium focus area” defined by the United States Geological Survey (USGS; Dicken et al. 2022). The lithium focus area is a broad, regionally delineated area with lithium deposits. These areas have documented concentrations of lithium in sediments or brines, and may be more likely to include projects that are developed into mine sites than other locations (Hammarstrom et al. 2020). Further, these areas are likely to receive greater pressures from the exploration process that often includes creating roads, drilling test wells, and collecting surface and subsurface ore samples.

Species records were obtained for the state of California from the California Natural Diversity Database (CNDDDB) and for Nevada from the Nevada Division of Natural Heritage (NDNH). Data were obtained from iNaturalist (research grade records), a citizen/community science database, in both states. Data were filtered and species occurrences older than 1990 were removed from the analyses because they may no longer be valid if they have not been updated during the past 30 years. We classified data into four categories: 1) inside the proposed lithium project area, including the 3.2 km buffer; 2) outside the proposed lithium project area beyond the 3.2 km buffer; 3) inside the focus area; and 4) outside the focus area (Figure 1).

We focused our analyses on the imperiled to vulnerable ranked

species, which globally are ranked as G1-G3 and T1-T2 and at the state level they are ranked as S1-S3. The globally ranked species are potentially at greater risk of extinction than S ranked species, because S ranked species may have a larger distribution occurring in another state or province. The S ranked species are often rare, occur at the edge of a species range, or they may be populations isolated from the main distribution of the species, and they may also include a G1-G3 species rank. The rankings are defined by NatureServe as:

G1: Critically Imperiled – at very high risk of extinction or elimination due to very restricted range, very few populations or occurrences, very steep declines, very severe threats, or other factors.

G2: Imperiled – at high risk of extinction or elimination due to restricted range, few populations or occurrences, steep declines, severe threats, or other factors.

G3: Vulnerable – at moderate risk of extinction or elimination due to a fairly restricted range, relatively few populations or occurrences, recent and widespread declines, threats, or other factors.

GX: Presumed Extinct – Not located despite intensive searches and virtually no likelihood of rediscovery.

GU: Unrankable – Unrankable due to lack of information or due to substantially conflicting information about status or trends.

T: Intraspecific Taxon (trinomial) – the status of infraspecific taxa (subspecies or varieties) are indicated by a “T-rank” following the species’ global rank. Rules for assigning T-ranks follow the same principles outlined above. Examples include a variety of a species like *Ivesia kingii* var. *eremica* (Ranked G4T1) or subspecies like *Euphilotes pallescens arenamontana* (Ranked G3?T1).

S1: Critically Imperiled – at very high risk of extirpation in the jurisdiction due to very restricted range, very few populations or occurrences, very steep declines, severe threats, or other factors.

S2: Imperiled – at high risk of extirpation in the jurisdiction due to restricted range, few populations or occurrences, steep declines, severe threats, or other factors.

S3: Vulnerable – at moderate risk of extirpation in the jurisdiction due to a fairly restricted range, relatively few populations or occurrences, recent and widespread declines, threats, or other factors.

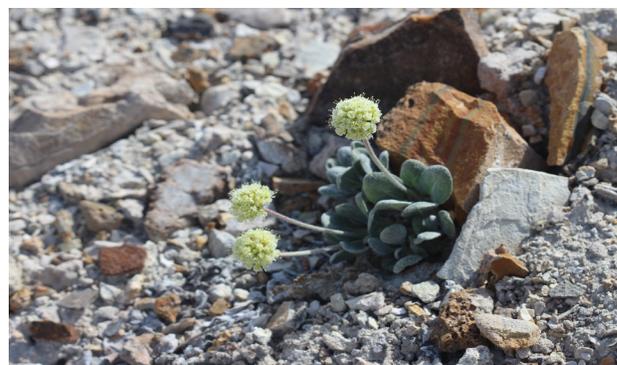
To provide consistent analyses, the lowest ranking was used to define a species. For example, a species with multiple rankings (e.g., G1G2) was considered at the lower ranking (G1) to provide the most conservative conservation estimate. In cases where a species had a T1 or T2 ranking, those rankings were prioritized over the species G ranking because the subspecies or variety warranted a higher conservation ranking. For example, *Hesperia uncas fulvapalla* is ranked as G4G5T1, but the subspecies is only known to occur in the Railroad Valley of central Nevada. Its T ranking was used in the analyses because otherwise the G ranking would lump the endemic subspecies with the larger species taxonomic grouping which is found broadly through western North America.

The location of some species occurrence records obtained from

iNaturalist were obscured by up to 30 km. When analyzing data in the potential lithium project areas we removed data with locations obscured by >1 km to avoid false positives/negatives on potential project locations. To analyze the species occurrences within the lithium focus area, we used all data including those with large location uncertainties, because the lithium focus area dataset is a regional dataset with broadly drawn boundaries.

We calculated the percentage of records occurring within proposed lithium projects and the lithium focus area to assess the overall potential impact of lithium mine development on individual species. The percentage of records within either the proposed lithium project or focus area provided an estimate of the proportion of a species range occurring within a project. The percentages of species recorded within the proposed lithium projects is not a direct assessment of the entirety of a species range, however it is a reasonable analog based on the best available data (Pearson et al. 2007).

The region where all proposed lithium projects were located is classified as desert and most of the lithium focus area in California and Nevada occurs in the desert regions. Because of the desert environment and importance of water resources to biodiversity, we analyzed wetland and spring information at each proposed lithium project. In California we used data from the National Wetland Inventory (NWI) and in Nevada we used data from the iGDE database (Saito et al. 2020). Spatial data from each dataset were overlaid and extracted at each proposed lithium project. Data were analyzed by calculating the amount of wetland area or number of springs within proposed lithium projects.



Amargosa niterwort (*Nitrophila mohavensis*) from the Amargosa Valley; *Tiehm's buckwheat* (*Eriogonum tiehmii*) from the Silver Peak Range, Nevada © Naomi Fraga



Fish Lake Valley in Esmeralda County, Nevada has several potential lithium projects © Michael Clifford/TNC

Results

We analyzed 15 proposed lithium projects in California and 40 proposed lithium projects in Nevada (Figure 1). Projects were not evenly distributed spatially, but were focused in specific regions of southern California near the Salton Sea, the playas or salars in western Nevada, and with ancient caldera landforms in northern and western Nevada. The proposed lithium projects in California comprised 77,702 acres (31,445 hectares) with 218,869 acres (88,563 ha) of buffer, while in Nevada proposed projects occurred on 309,339 acres (125,185 ha) with 724,593 acres (293,233 ha) of buffer. The lithium focus area occurred over 100,670 km² and 101,994 km² in California and Nevada, respectively. In California one proposed lithium project had a wetland feature, while in Nevada 36 proposed lithium projects had wetland features. There were three proposed projects in California with mapped springs located within the project boundary totaling 16 springs within the three projects. In Nevada, there were 27 proposed lithium projects with mapped springs located in the project boundary, and a total of 388 springs found within proposed projects. The total wetland area in proposed lithium projects was 98,626 acres (39,914 ha), of which over 99% of wetland area was found on proposed lithium projects in Nevada.

G1 and S1 Ranks - the critically imperiled species

There were 20 species recorded on proposed lithium projects that were ranked as G1/T1 or S1 and two species ranked GX and GU (Table 1). In California there were four G1/T1 ranked species which included *Agelaius tricolor*, *Cyprinodon macularius*, *Horkelia cuneata* var. *puberula*, and *Laterallus jamaicensis coturniculus*. In Nevada there were four G1 and three T1 ranked species. The GX and GU species both occurred in Nevada, which were *Pyrgulopsis*

ruinosa and *Anaxyrus* sp. 2, respectively. There were 10 S1 ranked species recorded in California and 11 S1 ranked species recorded in Nevada on proposed lithium projects. While any of the G1/T1 or S1 ranked species recorded within a proposed lithium project increases the risk of extirpation or extinction, seven of the G1/T1/S1 ranked species had 100% of their records occur within a proposed lithium project (Table 1). The seven species also included both of the GX and GU ranked species. Species with 100% of their records within a proposed lithium project or focus area could therefore become extinct if adverse impacts from lithium extraction occur. Of the species with 100% of their records observed within proposed lithium projects, only *Hulsea mexicana*, occurs in California, which captured the northernmost extent of its range; however, the species is known from a broader range outside of California and has historical records in California that were excluded from this analysis as they occurred prior to 1990. In Nevada, the six species were spread across the state, and were species with small ranges and small available habitat. For example, *Anaxyrus nevadensis* and *Anaxyrus* sp. 2, are endemic toad species occurring in wetlands in Railroad Valley and Fish Lake Valley, respectively. *Pyrgulopsis ruinosa* and *Pyrgulopsis lockensis* are freshwater snails found in only a few springs and each is located in a single valley.

Seventeen of the 20 species (85%) ranked G1/T1/S1 were dependent on wetlands for at least part of their life cycle, including both GX and GU ranked species. Further, five of the G1/T1/S1 ranked species with 100% of their records occurring within a proposed lithium project were wetland dependent.

California

In California, there were 669 species recorded in the lithium focus area. Of the species recorded in the lithium focus area, 367 were imperiled and vulnerable species ranked G1-G3 or T1 or T2 (Table 2; Appendix A). At the state level, 647 species were ranked S1-S3 in the lithium focus area. Nearly 70% of all recorded species in the lithium focus area were plants. Plants were also the most imperiled and vulnerable group, comprising 71% of broader taxonomic groups. Additionally, in California there were five special habitats with G3 ranks that were recorded in the lithium focus area which were Mojave Mixed Steppe, Mojave Yucca Scrub and Steppe, Valley Needlegrass Grassland, Water Birch Riparian Scrub, and Crucifixion Thorn Woodland (California Department of Fish and Wildlife 2023). Those special habitats also ranked between S1 and S3.

There were 61 species recorded within the proposed lithium project areas in California (Appendix B). There were 34 species ranked G1-G3 or T1-T2 and 58 species ranked S1-S3 recorded within proposed lithium projects (Figure 2). Within the proposed lithium projects, two species were ranked G1, *Cyprinodon macularius* and *Agelaius tricolor*, and 10 species were ranked S1 (Table 1). All the G1 species were also ranked S1 (it is common

in the NatureServe rankings to have overlap of G1 and S1 ranked species).

Nevada

A wide range of taxonomic groups were recorded in the lithium focus area of Nevada including insects, plants, mammals, and mollusks. Of the 304 species recorded in the lithium focus area, 190 species ranked G1-G3, including T ranked species, and there were two additional species ranked GU and GX. There were 267 species ranked S1-S3. Plants comprised more than half of the G-ranked species. There were 27 species ranked T1 and 15 species ranked T2. There were 116 species recorded in Nevada that had 100% of their records occur within the lithium focus area.

There were 43 species recorded within proposed lithium projects. There were 20 species ranked G1-G3, four species ranked T1-T2, one species ranked GX, one species ranked GU, and 38 species ranked S1-S3 recorded on proposed lithium projects. Plants comprised 33% of species recorded within proposed lithium projects (Table 2). While plants were also the most recorded species on the proposed lithium projects, mollusks and mammals were also present in relatively high proportions at >10% of species. Imperiled crustaceans and reptiles were not observed within the proposed lithium projects in Nevada.

Table 1. List of G1/T1 and S1 Ranked species recorded on lithium mine claims in California and Nevada. The GX and GU Ranked species are also included in the table.

Species	G rank, S rank	Percent in project	Wetland dependent	State
<i>Agelaius tricolor</i>	G1G2, S1S2	0.54%	Yes	CA
<i>Anaxyrus nevadensis</i>	G1, S1	100.00%	Yes	NV
<i>Anaxyrus</i> sp. 2	GU, S2	100.00%	Yes	NV
<i>Crenichthys nevadae</i>	G1, S1	16.67%	Yes	NV
<i>Cyprinodon macularius</i>	G1, S1	10.77%	Yes	CA
<i>Empidonax traillii extimus</i>	G5T2, S1	3.17%	Yes	CA
<i>Eriogonum tiehmii</i>	G1, S1	100.00%	No	NV
<i>Euphilotes pallescens arenamontana</i>	G3T1, S1	25.00%	No	NV
<i>Gelochelidon nilotica</i>	G5, S1	31.82%	Yes	CA
<i>Hesperia uncas fulvapalla</i>	G4G5T1, S1	100.00%	Yes	NV
<i>Horkelia cuneata</i> var. <i>puberula</i>	G4T1, S1	3.13%	No	CA
<i>Hulsea mexicana</i>	G3, S1	100.00%	No	CA
<i>Juga acutifilosa</i>	G2, S1	10.00%	Yes	NV
<i>Laterallus jamaicensis coturniculus</i>	G3G4T1, S1	0.51%	Yes	CA
<i>Lepidium integrifolium</i>	G2G3T2T3, S1	50.00%	Yes	NV
<i>Pelecanus erythrorhynchos</i>	G4, S1S2	0.29%	Yes	CA
<i>Penstemon albomarginatus</i>	G2, S1	4.55%	No	CA
<i>Pyrgulopsis lockensis</i>	G1, S2	100.00%	Yes	NV
<i>Pyrgulopsis ruinosa</i>	GX, S1	100.00%	Yes	NV
<i>Pyrgulopsis wongi</i>	G2, S1	25.00%	Yes	NV
<i>Rallus obsoletus yumanensis</i>	G3T3, S1S2	15.22%	Yes	CA
<i>Siphateles bicolor</i> ssp. 4	G4T1Q, S1	40.00%	Yes	NV

Table 2. Numbers of grouped species in California and Nevada on proposed lithium projects and the lithium focus area. Numbers in parentheses are the numbers of species with G1-G3 and T1-T2 Ranks.

Group	California		Nevada	
	Focus area	Proposed projects	Focus area	Proposed projects
Amphibians	15 (10)	2 (2)	7 (6)	3 (3)
Birds	65 (12)	25 (7)	37 (7)	4 (1)
Crustaceans	7 (7)	4 (4)	0 (0)	0 (0)
Fish	14 (11)	1 (1)	16 (13)	2 (2)
Insects	18 (17)	0 (0)	27 (20)	2 (2)
Mammals	49 (18)	4 (1)	36 (6)	16 (4)
Mollusks	15 (14)	0 (0)	44 (40)	5 (5)
Plants	462 (262)	21 (18)	124 (120)	10 (7)
Reptiles	24 (15)	4 (2)	14 (1)	1 (0)

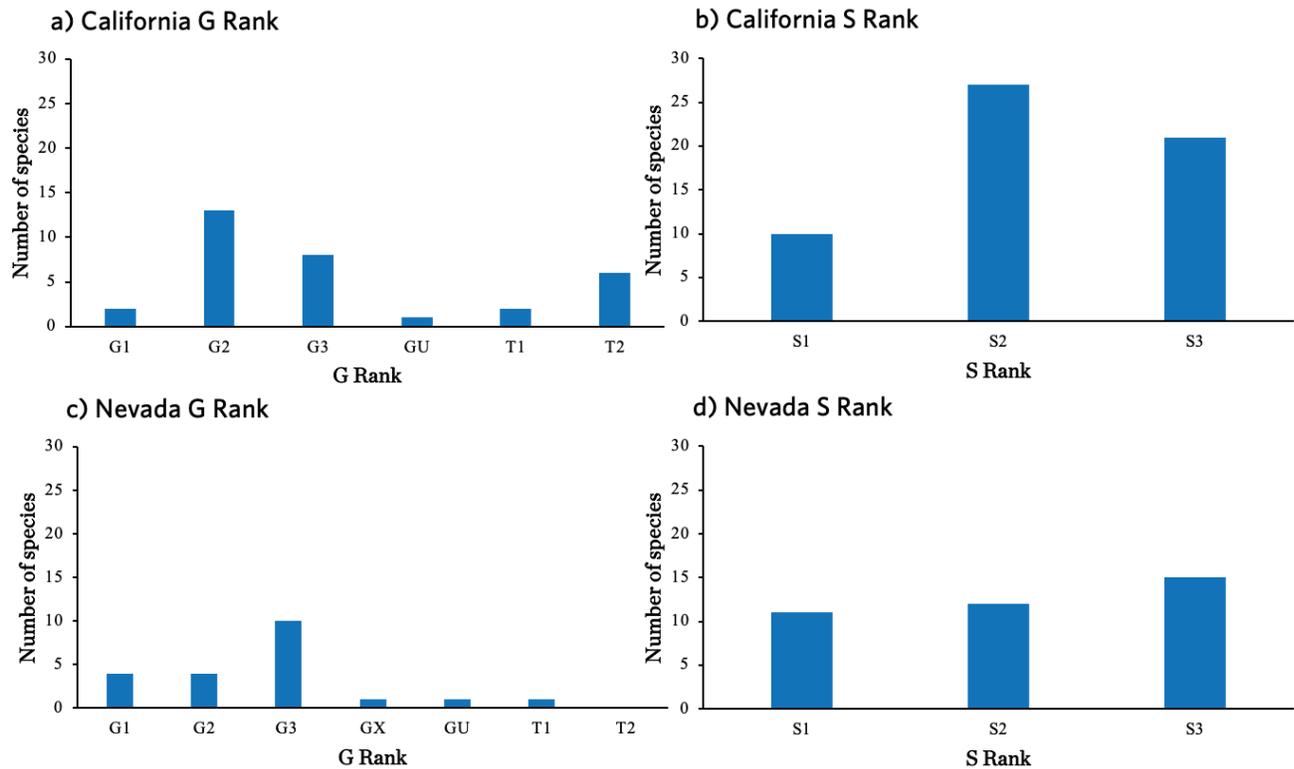


Figure 2. Number species recorded in proposed lithium projects in a) California G1-G3 and T1-T2 rank, b) California S1-S3 rank c) Nevada G1-G3 and T1-T2 rank, and d) Nevada S rank. Ranks of GX and GU are also included in G ranks.



The Amargosa River in southern California © Chip Carroon/TNC

Discussion

The development of proposed lithium projects in California and Nevada will negatively impact biodiversity if appropriate planning is not taken to avoid the most sensitive areas (e.g., Kiesecker et al. 2010). The 55 proposed projects in California and Nevada occur over a large area, broadly defined as desert with limited precipitation and surface water. Highlighting the importance of springs and wetlands to the biodiversity of the region, 17 of the 20 the most critically imperiled species recorded within proposed lithium projects were wetland dependent. More broadly, we identified 33 species in California with imperiled to vulnerable rankings on proposed lithium projects, while in Nevada we identified 24 species that were ranked imperiled to vulnerable. Even more broadly, we identified 367 species in the lithium focus area in California and 190 species in Nevada that were classified as globally imperiled or vulnerable.

We identified 20 critically imperiled species ranked G1/T1 and/or S1 recorded on proposed lithium projects. Further, 100% of the records for seven critically imperiled species occurred entirely within proposed lithium projects. These seven species face the greatest risk of extirpation or extinction if proposed lithium projects are developed. Several species, such as *Anaxyrus nevadensis*, *Eriogonum tiehmii*, and *Pyrgulopsis lockensis* are small ranged endemics and highlight the risk to such species (Primm et al. 2014). For example, *A. nevadensis* is an endemic species of toad occurring in isolated wetlands found in the Railroad Valley. *Eriogonum tiehmii* is a perennial plant that grows on less than

5 hectares in the Silver Peak Range, and was listed as federally endangered under the US Endangered Species Act in 2022 due to threats from mining. *Pyrgulopsis ruinosa*, an endemic spring snail located in Fish Lake Valley, Nevada also had 100% of its records within a proposed lithium project. However, *P. ruinosa* was presumed extinct (ranked GX) and was not observed between the early 1990s and 2020, despite efforts to locate the species (Springsnail Conservation Team 2020). *P. ruinosa* was finally observed in 2021 (E. Miskow, personal communication), and illustrates the difficulty in observing, monitoring, and tracking cryptic, small range endemic species. Regardless, project development covering the entire range of *P. ruinosa* will increase extinction risk due to habitat disturbance and/or mining-associated water pumping and use that could dry up its remaining habitat.

More broadly distributed species that have isolated or outlier populations such as *Hulsea mexicana* and *Hesperia uncas fulvapaella* have 100% of their records also occurring within a proposed lithium project. However, while these species are critically imperiled with rank S1, the existence of populations outside the states where they are rare and tracked as S1 species means that they are less likely to face extinction, and more likely to face local extirpation due to lithium project development. Other imperiled species with less than 100% of their range in a proposed lithium project still face increasing risks of extinction or extirpation as most of these species have small ranges, or small

population sizes, and even the loss of a small portion of habitat or population can negatively impact their long-term survival (Purvis et al. 2000; Primm et al. 2014; Staude et al. 2018). The addition of lithium extracting activities within their home range will greatly increase pressure on those species.

Many of the most imperiled species are endemic or outlier populations of species, which is a common global pattern of extinction (Enquist et al. 2019; Kraus et al. 2023). Endemic species and outlier populations such as *Anaxyrus nevadensis*, *Hulsea mexicana*, and *Hesperia uncas fulvapalla* play a pivotal role in biodiversity, having evolved within the unique ecosystems of localized conditions and often have specialized adaptations that enable them to survive in unique environmental conditions (Cantonati et al. 2020). The presence of endemic species and outlier populations can indicate past climate changes (Jansson 2003), and may provide insights into species range shifts under climate change. Endemic species and outlier populations also provide unique ecosystem services relative to cosmopolitan species (Gorman et al. 2014). The conservation of endemic species is important to reducing biodiversity loss. In many definitions (including in this report), endemic species are defined by political boundaries (Shipley and McGuire 2022; Kraus et al. 2023), and do not require bi-state policy measures to increase conservation – conservation can be achieved with intrastate policies. Without the need for multi-state conservation policy, endemic species should be quicker to protect because of the reduced interstate governmental agencies involved.

Of the 20 most critically imperiled species, 17 species are also dependent on wetlands for at least part of their life cycle (Table 1), highlighting the need for protections of wetlands. While there was a high proportion of wetland dependent species classified as critically imperiled and these species are at risk from surface development adjacent to springs and wetlands, our analyses do not include the potential impacts to wetland dependent species outside the proposed project boundary that may be impacted due to groundwater pumping from lithium extraction processes. Further, risks to groundwater dependent ecosystems differs based on the nearby type of lithium extraction (e.g., DLE, evaporative concentration, open pit, etc.). The hydrological framework and checklist developed by Saftner et al. (2023) will provide a path forward in considering hydrologic risks from potential impacts at the project level. However, the dependence of these species on water, which is often in the form of groundwater, highlights the need for detailed assessments of the hydrogeology in each basin with proposed lithium extraction, and larger comprehensive water management plans that consider the importance of water for natural ecosystems.

Even though DLE is expected to use a closed loop system for lithium extraction, more data are needed to understand potential negative impacts to wetlands and springs such as changes in spring discharge and temperature due to the changes in aquifer dynamics from pumping and reinjection (Kristmannsdottir and Armannsson 2003; Simmons et al. 2021).

We recognize there are data inequalities in the databases we used for locating species records (i.e., NDNH, CNDDDB, and iNaturalist). These inequalities occur in several ways, including

spatial inequalities where locations adjacent to large population centers have more data than rural or sparsely populated areas, which is particularly prominent in Nevada (Taylor 2014).

Another inequality is in the species identified, with more charismatic species groups like plants and birds being overrepresented relative to other more difficult to identify groups like insects. Taxonomic bias is well-documented in conservation science (Clark and May 2002). Furthermore, we recognize there are shortcomings to using only presence data, and that the absence of a species in the data does not imply absence on the landscape. We used the best available data to project the potential impacts to biodiversity from an emerging land use and urge increased biological collections in the lithium focus area so that biodiversity is not impacted due to a lack of data. Many species remain undiscovered, poorly studied, or have only recently been described. This leaves gaps in our understanding of their ecological roles, distribution, vulnerabilities, and population trends. The paucity of species information exacerbates the challenges of conservation, creating significant obstacles to understanding which species are affected or lost (e.g., Primm et al. 2014). There are still taxonomic uncertainties in many species and the lack of comprehensive data hampers our ability to accurately assess the status of various ecosystems and identify species at risk. In the lithium focus areas of California and Nevada combined, there were 138 species ranked as GU or T1-T2. The taxonomic uncertainties in many of the species recorded in the lithium focus area or in proposed lithium projects further challenges conservation. Policy protections from government agencies may not apply to these taxonomically ambiguous or undescribed species as they do not fall under governmental protections. However, citizen/community science data has the potential to rapidly alter our understanding of species and their distributions through discoveries of new populations, range boundaries (Kohler et al. 2023), and new species by making images and locations broadly available to all (e.g., Amezcuita et al. 2013; Jain et al. 2022).

Addressing global climate change is critically important for ecosystems and people, but climate change solutions must be balanced with the conservation of biodiversity (Wu et al. 2023). High biodiversity maintains ecosystem function, and typically sequesters more carbon than degraded systems experiencing extirpation. Extinction of cryptic species may signal weakening of ecosystem function and provide early indications of poorly functioning ecosystems. The loss of biodiversity significantly decreases ecosystem resiliency and ecosystem services that both humans and nature depend on for survival (Cardinale et al. 2012). While the ongoing renewable energy transition is a key component of climate change action, the deployment of technologies and resource extraction activities focused on furthering the transition needs to be planned so that projects avoid the most sensitive species areas, and consume the least amount of water, especially in arid regions.

References

- Amézquita A, R Márquez, R Medina, D Mejía-Vargas, TR Kahn, G Suárez, L Mazariegos. 2013. A new species of Andean poison frog, *Andinobates* (Anura: Dendrobatidae), from the northwestern Andes of Colombia. *Zootaxa* 3620: 163-178.
- Bradley DC, LL Stillings, BW Jaskula, L Munk, AD McCauley. 2017. Lithium: Chapter K of critical mineral resources of the United States—economic and environmental geology and prospects for future supply. Eds KJ Schulz, JH DeYoung, Jr., RR Seal II, DC Bradley. US Geological Survey Professional Paper 1802-K.
- California Department of Fish and Wildlife. (2023) California Natural Diversity Database (CNDDDB) – Government version downloaded April 30, 2023.
- Cantonati M, S Poikane, CM Pringle, LE Stevens, E Turak, J Heino, JS Richardson, R Bolpagni, A Borrini, N Cid, M Ctvrtlikova, DM Galassi, M Hajek, I Hawes, Z Levkov, L Naselli-Flores, AA Saber, M Di Cicco, B Fiasca, PB Hamilton, J Kubecka, S Segadelli, P Znachor. 2020. Characteristics, main impacts, and stewardship of natural and artificial freshwater environments: Consequences for biodiversity conservation. *Water* 12:260. Doi: 10.3390/w12010260.
- Cardinale BJ, JE Duffy, A Gonzalez, DU Hooper, C Perrings, P Venail, A Narwani, GM Mace, D Tilman, DA Wardle, AP Kinzig, GC Daily, M Loreau, JB Grace, A Larigauderie, DS Srivastava, S Naeem. 2012. Biodiversity loss and its impact on humanity. *Nature* 486:59-67.
- Ceballos G, PR Ehrlich, AD Barnosky, A García, RM Pringle, TM Palmer. 2015. Accelerated modern human-induced species losses: entering the sixth mass extinction. *Science Advances* 1:e1400253.
- Chapin III SF, ES Zavaleta, VT Eviner, RL Naylor, PM Vitousek, HL Reynolds, DU Hooper, S Lavorel, OE Sala, SE Hobbie, MC Mack, S Díaz. 2000. Consequences of changing biodiversity. *Nature* 405:234-242.
- Chen I-C, JK Hill, R Ohlemüller, DB Roy, CD Thomas. 2011. Rapid Range Shifts of Species Associated with High Levels of Climate Warming. *Science* 333:1024-1026.
- Clark JA, RM May. 2002. Taxonomic bias in conservation research. *Science* 297:191-192.
- Dashiell S, M Buckley, D Mulvaney. 2019. Green light study: economic and conservation benefits of low-impact solar siting in California. *ECONorthwest and The Nature Conservancy*, pp. 45.
- Davis J, A Pavlova, R Thompson, P Sunnucks. 2013. Evolutionary refugia and ecological refuges: key concepts for conserving Australian arid zone freshwater biodiversity under climate change. *Global Change Biology* 19:1970-1984.
- Diaz S, J Settele, ES Brondízio, HT Ngo, J Agard, A Arneeth, P Balvanera, KA Brauman, SHM Butchart, KMA Chan, LA Garibaldi, K Ichii, J Liu, SM Subramanian, GF Midgley, Patricia Miloslavich, Z Molnár, D Obura, A Pfaff, S Polasky, A Purvis, J Razzaque, B Reyers, RR Chowdhury, Y-J Shin, I Visseren-Hamakers, KJ Willis, CN Zayas. 2019. Pervasive human-driven decline of life on Earth points to the need for transformative change. *Science* 366:6471
- Dicken CL, LG Woodruff, JM Hammarstrom, KE Crocker. 2022. GIS, supplemental data table, and references for focus areas of potential domestic resources of critical minerals and related commodities in the United States and Puerto Rico: U.S. Geological Survey data: <https://doi.org/10.5066/P9DIZ9N8>.
- Flexer V, CF Baspineiro, CI Galli. 2018. Lithium recovery from brines: A vital raw material for green energies with a potential environmental impact in its mining and processing. *Science of the Total Environment* 639:1188-1204.
- Gorman CE, BM Potts, JA Schweitzer, JK Bailey. 2014. Shifts in species interactions due to the evolution of functional differences between endemics and non-endemics: an endemic syndrome hypothesis. *PLoS ONE* 9: e111190. doi:10.1371/journal.pone.0111190.
- Haddad AZ, L Hackl, B Akuzum, G Pohlman, J-F Magnan, R Kostecki. 2023. How to make lithium extraction cleaner, faster and cheaper - in six steps. *Nature* 616:245-248.
- Hammarstrom J, C Dicken, W Day, A Hofstra, B Drenth, A Shah, A McCafferty, L Woodruff, N Foley, D Ponce, T Frost, L Stillings. 2020. Focus areas for data acquisition for potential domestic resources of 11 critical minerals in the conterminous United States, Hawaii, and Puerto Rico—Aluminum, cobalt, graphite, lithium, niobium, platinum-group elements, rare earth elements, tantalum, tin,

- titanium, and tungsten (ver. 1.1, July 2022), chap. B of U.S. Geological Survey, Focus areas for data acquisition for potential domestic sources of critical minerals: U.S. Geological Survey Open-File Report 2019-1023, 67 p., <https://doi.org/10.3133/ofr20191023B>.
- Herrington R. 2021. Mining our green future. *Nature Reviews Materials* 6:456-458.
- Hise C, B Obermeyer, M Ahlering, J Wilkinson, J Fargione. 2022. Site Wind Right: Identifying Low-Impact Wind Development Areas in the Central United States. *Land* 11:462. <https://doi.org/10.3390/land11040462>.
- Jain P, H Forbes, LA Esposito. 2022. Two new alkali-sink specialist species of *Paruroctonus* Werner 1934 (Scorpiones, Vaejovidae) from central California. *ZooKeys* 1117:139-188.
- Jansson R. 2003. Global patterns in endemism explained by past climatic change. *Proceedings of the Royal Society B* 270:583-590.
- Kohler DB, BT Hamilton, DE Dittmer, AS Whiting. 2023. Citizen Science in Action: An Updated Distribution for *Lampropeltis pyromelana*. *Western North American Naturalist* 83:165-175.
- Kristmannsdottir H and H Armannsson. 2003. Environmental aspects of geothermal energy utilization. *Geothermics* 32:451-461.
- IPCC. 2018. Global Warming of 1.5°C. An IPCC Special Report on the impacts of global warming of 1.5°C above pre-industrial levels and related global greenhouse gas emission pathways, in the context of strengthening the global response to the threat of climate change, sustainable development, and efforts to eradicate poverty [Masson-Delmotte, V, P Zhai, H-O Pörtner, D Roberts, J Skea, PR Shukla, A Pirani, W Moufouma-Okia, C Péan, R Pidcock, S Connors, JBR Matthews, Y Chen, X. Zhou, MI Gomis, E Lonnoy, T Maycock, M Tignor, and T Waterfield (eds.)]. Cambridge University Press, Cambridge, UK and New York, NY, USA, 616 pp., doi:10.1017/9781009157940.
- Jowitt SM and BA McNulty. 2021. Battery and energy metals: future drivers of the minerals industry? *SEG Discovery* 127:11-18.
- Kiesecker JM, H Copeland, A Pocewicz, B McKenney. 2010. Development by design: blending landscape level planning with the mitigation hierarchy. *Frontiers in the Environment and Ecology* 8:261-266.
- Kraus D, A Enns, A Hebb, S Murphy, DAR Drake, B Bennett. 2023. Prioritizing nationally endemic species for conservation. *Conservation Science and Practice* 5:e12845.
- Obermeyer B, R Manes, J Kiesecker, J Fargione, K Sochi. 2011. Development by design: mitigating wind development's impacts on wildlife in Kansas. *PLoS ONE* 6:e26698. doi:10.1371/journal.pone.0026698.
- Park DS, RK Peet, M Pillet, JM Serra-Diaz, B Sandel, M Schildhauer, I Šímová, C Violle, JJ Wieringa, SK Wisser, L Hannah, J-C Svenning, BJ McGill. 2019. The commonness of rarity: global and future distribution of rarity across land plants. *Science Advances* 5:eaaz0414.
- Parker SS, A Zdon, WT Christian, BS Cohen, M Palacios Mejia, NS Fraga, EE Curd, K Edalati, MA Renshaw. 2021. Conservation of Mojave Desert springs: status, threats, and policy opportunities. *Biodiversity and Conservation* 30:311-327. doi:10.1007/s10531-020-02090-7.
- Parker SS, BS Franklin, A Williams, BS Cohen, MJ Clifford, MM Rohde. 2022. Potential Lithium extraction in the United States: environmental, economic, and policy implications. *The Nature Conservancy*: <https://www.scienceforconservation.org/products/lithium>
- Parker SS, MJ Clifford, BS Cohen. 2024. Potential impacts of proposed lithium extraction on biodiversity and conservation in the contiguous United States. *Science of the Total Environment*. doi:10.1016/j.scitotenv.2023.168639.
- Pearson RG, CJ Raxworthy, M Nakamura, AT Peterson. 2007. Predicting species distributions from small numbers of occurrence records: a test case using cryptic geckos in Madagascar. *Journal of Biogeography* 34:102-117.
- Pörtner H-O, RJ Scholes, A Arneeth, DKA Barnes, MT Burrows, SE Diamond, CM Duarte, W Kiessling, P Leadley, S Managi, P McElwee, G Midgley, HT Ngo, D Obura, U Pascual, M Sankaran, YJ Shin, AL Val. 2023. Overcoming the coupled climate and biodiversity crises and their societal impacts. *Science* 380:eabl4881. doi:10.1126/science.abl4881.
- Pounds JA, MR Bustamante, LA Coloma, JA Consuegra, MPL Fogden, PN Foster, E La Marca, KL Masters, A Merino-Viteri, R Puschendorf, SR Ron, GA Sanchez-Azofeifa, CJ Still, BE Young. 2006. Widespread amphibian extinctions from epidemic disease driven by global warming. *Nature* 439:161-167.

- Powers RP and W Jetz. 2019. Global habitat loss and extinction risk of terrestrial vertebrates under future land-use-change scenarios. *Nature Climate Change* 9: 323-329.
- Primm SL, CN Jenkins, R Abell, TM Brooks, JL Gittleman, LN Joppa, PH Raven, CM Roberts, JO Sexton. 2014. The biodiversity of species and their rates of extinction, distribution, and protection. *Science* 344:1246752.
- Purvis A, JL Gittleman, G Cowlishaw, GM Mace. 2000. Predicting extinction risk in declining species. *Proceedings of the Royal Society B: Biological Sciences* 267:1947-1952.
- Randall JM, SS Parker, J Moore, B Cohen, L Crane, B Christian, D Cameron, J MacKenzie, K Klausmeyer, S Morrison. 2010. Mojave Desert Ecoregional Assessment. Unpublished Report. The Nature Conservancy, San Francisco, California. 106 pp + appendices. <https://www.scienceforconservation.org/products/mojave-desert-ecoregional-assessment>
- Saftner D, K Heintz, R Hershey. 2023. Identifying potential hydrologic impacts of lithium extraction in Nevada. Publication No. 41297. Reno: Desert Research Institute. https://www.groundwaterresourcehub.org/content/dam/tnc/nature/en/documents/groundwater-resource-hub/saftner23_hydro_impacts_lithium.pdf
- Saito L, S Byer, K Badik, K McGwire, L Provencher, B Minor. 2020. Mapping indicators of groundwater dependent ecosystems in Nevada: Important resources for a water-limited state. *Journal of Nevada Water Resources Association*, Winter 2020: 48-72. DOI: 10.22542/jnwra/2020/1/3.
- Saito L, S Byer, K Badik, L Provencher, D McEvoy. 2022. Stressor and Threat Assessment of Nevada Groundwater Dependent Ecosystems. Reno: The Nature Conservancy. Available at <https://www.groundwaterresourcehub.org/where-we-work/nevada/nevada-gde-stressor-threat/>
- Sevico BK, SH Ali, M Bazilian, B Radley, B Nemery, J Okatz, D Mulvaney. 2020. Sustainable minerals and metals for a low-carbon future. *Science* 367:30-33.
- Shiple BR and JL McGuire. 2022. Interpreting and integrating multiple endemism metrics to identify hotspots for conservation priorities. *Biological Conservation* 265:109403.
- Simmons SF, RG Allis, SM Kirby, JN Moore, TP Fischer. 2021. Interpretation of hydrothermal conditions, production-injection induced effects, and evidence for enhanced geothermal system-type heat exchange in response to >30 years of production at Roosevelt Hot Springs, Utah, USA: *Geosphere* 17:1997-2026.
- Sinervo B, F Méndez-de-la-Cruz, DB Miles, B Heulin, E Bastiaans, M Villagrán-Santa Cruz, R Lara-Resendiz, N Martínez-Méndez, ML Calderón-Espinosa, RN Meza-Lázaro, H Gadsden, LJ Avila, M Morando, IJ De la Riva, PV Sepulveda, CFD Rocha, N Ibagüen-goytía, CA Puntriano, M Massot, V Lepetz, TA Oksanen, DG Chapple, AM. Bauer, WR Branch, J Clobert, JW Sites Jr. 2010. Erosion of lizard diversity by climate change and altered thermal niches. *Science* 328:894-899.
- Sonter LJ, MC Dade, JEM Watson, RK Valenta. 2020. Renewable energy production will exacerbate mining threats to biodiversity. *Nature Communications* 11:4174.
- Sonter LJ, M Marona, JW Bull, S Giljum, S Luckeneder, V Mause, E McDonald-Madden, SA Northey, LE Sánchez, R Valenta, P Visconti, TT Werner, JEM Watson. 2023. How to fuel an energy transition with ecologically responsible mining. *Proceedings of the National Academy of Sciences* 120:e2307006120.
- Springnail Conservation Team. 2020. Conservation strategy for springsnails in Nevada and Utah, Version 1.0. Nevada Department of Wildlife, Reno, and Utah Division of Wildlife Resources, Salt Lake City.
- Stade IR, LM Navarro, HM Pereira. 2020. Range size predicts the risk of local extinction from habitat loss. *Global Ecology and Biogeography* 29:16-25.
- Taylor DW. 2014. Large data inequalities in herbarium specimen density in the western United States. *Phytoneuron* 53:1-8.
- USGS. 2023. Minerals commodity summary: Lithium. National Minerals Information Center, US Geological Survey.
- Vera ML, WR Torres, CI Galli, A Chagnes, V Flexer. 2023. Environmental impact of direct lithium extraction from brines. *Nature*

Reviews Earth and Environment 4:149-165.

von Jeetze PJ, I Weindl, JA Johnson, P Borrelli, P Panagos, EJ Molina Bacca, K Karstens, F Humpenöder, JP Dietrich, S Minoli, C Müller, H Lotze-Campen, A Popp. 2023. Projected landscape-scale repercussions of global action for climate and biodiversity protection. *Nature Communications* 14:2515 <https://doi.org/10.1038/s41467-023-38043-1>.

Wu GC, RA Jones, E Leslie, JH Williams, A Pascale, E Brand, SS Parker, BS Cohen, JE Fargione, J Souder, M Batres, MG Gleason, MH Schindel, CK Stanley. 2023. Minimizing habitat conflicts in meeting net-zero energy targets in the western United States. *Proceedings of the National Academy of Sciences* 120: e2204098120.

CLOCKWISE FROM LEFT Killdeer (*Charadrius vociferus*) in Nevada © Michael Swink; White-margin beardtongue (*Penstemon albomarginatus*) in Nevada; Tecopa birdsbeak (*Chloropyron tecopense*) in Fish Lake Valley, Nevada; Silver Peak Range © Naomi Fraga BACK PAGE Tecopa, California © Chip Carroon/TNC



Appendix A. List of Species with G rank and S rank within the lithium focus area, with percent of total occurrences. Some species were recorded in both California and Nevada. The first number corresponds to the percentage in California and second number listed for Nevada.

Species	Percent in Focus area	G Rank	S Rank	State
<i>Abronia villosa</i> var. <i>aurita</i>	27.69%	G5T2?	S2	CA
<i>Abutilon parvulum</i>	50.00%	G5	S2S3	CA
<i>Acanthoscyphus parishii</i> var. <i>cienegeensis</i>	47.83%	G4?T2	S2	CA
<i>Acanthoscyphus parishii</i> var. <i>goodmaniana</i>	100.00%	G4?T1	S1	CA
<i>Accipiter cooperii</i>	1.14%	G5	S4	CA
<i>Accipiter gentilis</i>	4.21%, 31.08%	G5	S3, S3	CA, NV
<i>Acleisanthes nevadensis</i>	41.67%	G4?	S1	CA
<i>Acmispon argyraeus</i> var. <i>multicaulis</i>	33.33%	G4?T2	S2	CA
<i>Acmispon haydonii</i>	50.00%	G3	S3	CA
<i>Aegialia crescenta</i>	100.00%	G1	S1	NV
<i>Agastache cusickii</i>	100.00%	G3G4	S2	NV
<i>Agave utahensis</i> var. <i>eborispina</i>	100.00%	G4T3	S2	CA
<i>Agelaius tricolor</i>	2.48%	G1G2	S1S2	CA
<i>Ageratina herbacea</i>	100.00%	G5	S3	CA
<i>Agrilus harenus</i>	100.00%	G1G2	S1S2	CA
<i>Agrostis humilis</i>	35.29%	G4Q	S2	CA
<i>Aimophila ruficeps canescens</i>	2.55%	G5T3	S3	CA
<i>Aliciella ripleyi</i>	100.00%	G3	S2	CA
<i>Aliciella triodon</i>	57.14%	G5	S2	CA
<i>Allium atrorubens</i> var. <i>atorrubens</i>	42.86%	G4T4	S2	CA
<i>Allium marvinii</i>	2.27%	G1	S1	CA
<i>Allium nevadense</i>	26.32%	G4	S3	CA
<i>Allium shevockii</i>	83.33%	G2	S2	CA
<i>Almutaster pauciflorus</i>	100.00%	G4	S1S2	CA
<i>Ambystoma macrodactylum sigillatum</i>	1.52%	G5T4	S3	CA
<i>Anaxyrus californicus</i>	12.31%	G2G3	S2S3	CA
<i>Anaxyrus canorus</i>	10.89%	G2G3	S2S3	CA
<i>Anaxyrus monfontanus</i>	100.00%	G1	S1	NV
<i>Anaxyrus nelsoni</i>	100.00%	G2	S2	NV
<i>Anaxyrus nevadensis</i>	100.00%	G1	S1	NV
<i>Anaxyrus</i> sp. 2	100.00%	GU	S2	NV
<i>Androstephium breviflorum</i>	84.35%	G4	S2?	CA
<i>Anniella campi</i>	37.50%	G1G2	S1S2	CA
<i>Anniella pulchra</i>	12.71%	G3	S3	CA
<i>Anniella stebbinsi</i>	5.00%	G3	S3	CA

<i>Anodonta californiensis</i>	25%, 40%	G3Q	S2?, S1	CA, NV
<i>Antennaria marginata</i>	50.00%	G4G5	S1	CA
<i>Antigone canadensis tabida</i>	0.47%, 20%	G5T5	S2, S2BM	CA, NV
<i>Antrozous pallidus</i>	18.7%, 31.7%	G4	S3, S3	CA, NV
<i>Aplodontia rufa</i>	57.14%	G5	S1	NV
<i>Aplodontia rufa californica</i>	6.52%	G5T3T4	S2S3	CA
<i>Aquila chrysaetos</i>	22.5%, 16.7%	G5	S3, S4	CA, NV
<i>Arabis rigidissima</i> var. <i>demota</i>	50.00%	G3T3Q	S2	NV
<i>Arctomecon merriamii</i>	83.3%, 17.9%	G3	S3, S3	CA, NV
<i>Arctostaphylos glandulosa</i> ssp. <i>gabrielensis</i>	16.13%	G5T3	S3	CA
<i>Ardea alba</i>	7.41%	G5	S4	CA
<i>Ardea herodias</i>	3.95%	G5	S4	CA
<i>Arenaria lanuginosa</i> var. <i>saxosa</i>	7.69%	G5T5	S2	CA
<i>Argynnis nokomis apacheana</i>	15.38%	G3T3	S3	NV
<i>Arizona elegans</i>	14.29%	G5	S4	NV
<i>Arizona elegans occidentalis</i>	10.09%	G5T2	S2	CA
<i>Artemia monica</i>	100.00%	G3	S3	CA
<i>Artemisia tripartita</i> ssp. <i>tripartita</i>	75.00%	G5T4T5	S2	CA
<i>Artemisiospiza belli belli</i>	1.64%	G5T2T3	S3	CA
<i>Asclepias eastwoodiana</i>	18.18%	G2	S2S3	NV
<i>Asclepias nyctaginifolia</i>	23.44%	G4?	S2	CA
<i>Asio flammeus</i>	30.00%	G5	S2	NV
<i>Asio otus</i>	26.83%, 30.77%	G5	S3?, S3	CA, NV
<i>Aspidoscelis hyperythra</i>	0.39%	G5	S2S3	CA
<i>Aspidoscelis tigris stejnegeri</i>	1.12%	G5T5	S3	CA
<i>Assiminea infima</i>	100.00%	G1	S1	CA
<i>Astragalus albens</i>	100.00%	G1	S1	CA
<i>Astragalus allochrous</i> var. <i>playanus</i>	100.00%	G4T4	S2	CA
<i>Astragalus argophyllus</i> var. <i>argophyllus</i>	100.00%	G5T4	S2	CA
<i>Astragalus atratus</i> var. <i>mensanus</i>	77.78%	G4G5T2	S2	CA
<i>Astragalus austiniiae</i>	23.53%, 100%	G2G3	S2S3, S1	CA, NV
<i>Astragalus beatleyae</i>	100.00%	G2	S2	NV
<i>Astragalus bernardinus</i>	95.00%	G3	S3	CA
<i>Astragalus callithrix</i>	100.00%	G3	S3	NV
<i>Astragalus calycosus</i> var. <i>monophyllidius</i>	50.00%	G5T2Q	S3	NV
<i>Astragalus cimae</i> var. <i>cimae</i>	50.00%	G3T2T3	S2?	CA
<i>Astragalus cimae</i> var. <i>sufflatus</i>	33.33%	G3T3	S3	CA
<i>Astragalus ertterae</i>	33.33%	G2	S2	CA
<i>Astragalus funereus</i>	76.92%	G2	S2	NV

<i>Astragalus gilmanii</i>	100.00%	G2	S2	CA
<i>Astragalus insularis</i> var. <i>harwoodii</i>	47.57%	G5T4	S2	CA
<i>Astragalus jaegerianus</i>	100.00%	G2	S2	CA
<i>Astragalus johannis-howellii</i>	70%, 40%	G2	S1, S2	CA, NV
<i>Astragalus kentrophyta</i> var. <i>ungulatus</i>	100.00%	G5T3T4	S1	CA
<i>Astragalus lemmonii</i>	42.86%	G2	S2	CA
<i>Astragalus lentiginosus</i> var. <i>antoni</i>	60.00%	G5T2	S2	CA
<i>Astragalus lentiginosus</i> var. <i>coachellae</i>	100.00%	G5T1	S1	CA
<i>Astragalus lentiginosus</i> var. <i>latus</i>	23.08%	G5T2	S2	NV
<i>Astragalus lentiginosus</i> var. <i>piscinensis</i>	100.00%	G5T1	S1	CA
<i>Astragalus lentiginosus</i> var. <i>scorpionis</i>	37.50%	G5T3T4	S3?	NV
<i>Astragalus lentiginosus</i> var. <i>sesquimetalis</i>	100%, 100%	G5T1	S1, S1	CA, NV
<i>Astragalus lentiginosus</i> var. <i>sierrae</i>	31.58%	G5T2	S2	CA
<i>Astragalus leucolobus</i>	57.39%	G2	S2	CA
<i>Astragalus magdalenae</i> var. <i>peirsonii</i>	100.00%	G3G4T1	S1	CA
<i>Astragalus mohavensis</i> var. <i>hemigyryus</i>	100.00%	G3G4T2T3	S1	CA
<i>Astragalus mohavensis</i> var. <i>mohavensis</i>	4.00%	G3G4T3T4	S2S3	NV
<i>Astragalus monoensis</i>	96.15%	G2	S2	CA
<i>Astragalus nyensis</i>	6.25%	G3	S3	NV
<i>Astragalus oophorus</i> var. <i>clokeyanus</i>	31.03%	G4T2	S2	NV
<i>Astragalus phoenix</i>	100.00%	G2	S1	NV
<i>Astragalus platytropis</i>	28.57%	G5	S3	CA
<i>Astragalus porrectus</i>	40.00%	G3?	S3?	NV
<i>Astragalus preussii</i> var. <i>laxiflorus</i>	100.00%	G4T2	S1	CA
<i>Astragalus pseudiodanthus</i>	100%, 71%	G2Q	S2, S2	CA, NV
<i>Astragalus pterocarpus</i>	62.50%	G3	S3	NV
<i>Astragalus pulsiferae</i> var. <i>coronensis</i>	100.00%	G4T3	S1	NV
<i>Astragalus ravenii</i>	100.00%	G2	S2	CA
<i>Astragalus sabulonum</i>	12.5%, 33.3%	G4G5	S2, N/A	CA, NV
<i>Astragalus serenoii</i> var. <i>shockleyi</i>	71.43%	G4T3	S3	CA
<i>Astragalus serenoii</i> var. <i>sordescens</i>	100.00%	G4T2?	S2	NV
<i>Astragalus tidestromii</i>	56.25%	G4	S2	CA
<i>Astragalus tiehmii</i>	100.00%	G3	S2	NV
<i>Astragalus toquimanus</i>	33.33%	G2	S2	NV
<i>Astragalus tricarinatus</i>	98.04%	G2	S2	CA
<i>Astragalus yoder-williamsii</i>	50.00%	G3	S1	NV
<i>Astrolepis cochisensis</i> ssp. <i>cochisensis</i>	93.33%	G5?T4	S2	CA
<i>Athene cunicularia</i>	30%, 60%	G4	S3, NA	CA, NV
<i>Athene cunicularia hypugaea</i>	8.79%	G4T4	S3B	NV
<i>Atriplex argentea</i> var. <i>longitrichoma</i>	25.00%	G5T2	S2	CA
<i>Auriparus flaviceps</i>	19.30%	G5	S3	NV

<i>Ayenia compacta</i>	18.97%	G4	S3	CA
<i>Bahia neomexicana</i>	87.50%	G5	S2S3	CA
<i>Batrachoseps campi</i>	90.91%	G3	S3	CA
<i>Batrachoseps major aridus</i>	100.00%	G4T1	S1	CA
<i>Batrachoseps robustus</i>	53.85%	G3	S3	CA
<i>Berberis fremontii</i>	45.45%	G5	S3	CA
<i>Blepharidachne kingii</i>	25%, 66%	G4	S2, N/A	CA, NV
<i>Boechera bodiensis</i>	65%, 8.33%	G3	S3, S2	CA, NV
<i>Boechera cobrensis</i>	33.33%	G5	S3	CA
<i>Boechera dispar</i>	89%, 100%	G3	S3, S1S2	CA, NV
<i>Boechera johnstonii</i>	75.00%	G1	S1	CA
<i>Boechera lincolnensis</i>	100.00%	G4G5	S3	CA
<i>Boechera ophira</i>	36.36%	G1G2	S1	NV
<i>Boechera parishii</i>	58.06%	G2	S2	CA
<i>Boechera peirsonii</i>	50.00%	G1	S1	CA
<i>Boechera pendulina</i>	28.57%	G5	S2	CA
<i>Boechera pinzliae</i>	100.00%	G2	S1	CA
<i>Boechera rectissima x Boechera retrofracta</i>	42.86%	GNA	S1	NV
<i>Boechera shockleyi</i>	69.56%, 35.29%	G3	S2, S3	CA, NV
<i>Boechera tiehmii</i>	100%, 100%	G3	S3, S1	CA, NV
<i>Boechera tularensis</i>	21.43%	G3	S3	CA
<i>Bombus crotchii</i>	11.03%	G3G4	S1S2	CA
<i>Bombus morrisoni</i>	58%, 33%	G4G5	S1S2, N/A	CA, NV
<i>Bombus occidentalis</i>	5.00%	G2G3	S1	CA
<i>Botrychium ascendens</i>	11.76%	G3G4	S2	CA
<i>Botrychium crenulatum</i>	10.67%	G4	S3	CA
<i>Botrychium minganense</i>	1.28%	G4G5	S3	CA
<i>Brachylagus idahoensis</i>	70%, 31%	G4	S3, S3	CA, NV
<i>Branchinecta lynchi</i>	0.13%	G3	S3	CA
<i>Branta hutchinsii leucopareia</i>	4.35%	G5T3	S3	CA
<i>Bursera microphylla</i>	6.45%	G4	S2	CA
<i>Buteo regalis</i>	21%, 40%	G4	S3S4; S3B,S4N	CA, NV
<i>Buteo swainsoni</i>	2.50%, 79%	G5	S3, S3B	CA, NV
<i>Calliandra eriophylla</i>	80.56%	G5	S3	CA
<i>Calochortus clavatus var. gracilis</i>	0.74%	G4T2T3	S2S3	CA
<i>Calochortus excavatus</i>	100.00%	G2	S2	CA
<i>Calochortus leichtlinii</i>	100.00%	G4	S3	NV
<i>Calochortus palmeri var. munzii</i>	11.36%	G3T3	S3	CA
<i>Calochortus palmeri var. palmeri</i>	44.68%	G3T2	S2	CA

<i>Calochortus plummerae</i>	1.95%	G4	S4	CA
<i>Calochortus striatus</i>	93%, 56%	G3?	S2S3, S1	CA, NV
<i>Calyptridium pygmaeum</i>	66.67%	G1G2	S1S2	CA
<i>Camissonia integrifolia</i>	25.00%	G2	S2	CA
<i>Carex davyi</i>	28.57%	G3	S3	CA
<i>Carex duriuscula</i>	40.00%	G5	S2	CA
<i>Carex limosa</i>	6.25%	G5	S3	CA
<i>Carex occidentalis</i>	20.00%	G4	S3	CA
<i>Carex petasata</i>	3.08%	G5	S3	CA
<i>Carex praticola</i>	16.67%	G5	S2	CA
<i>Carex scirpoidea</i> ssp. <i>pseudoscirpoidea</i>	66.67%	G5T4	S2	CA
<i>Carex tiogana</i>	50.00%	G2Q	S1	CA
<i>Carex vallicola</i>	16.67%	G5	S2	CA
<i>Carnegiea gigantea</i>	8.00%	G5	S1	CA
<i>Castela emoryi</i>	92.00%	G3G4	S2S3	CA
<i>Castilleja cinerea</i>	28.26%	G1G2	S1S2	CA
<i>Castilleja gleasoni</i>	14.29%	G2	S2	CA
<i>Castilleja lasiorhyncha</i>	71.43%	G2?	S2?	CA
<i>Castilleja salsuginosa</i>	66.67%	G1	S1	NV
<i>Catostomus fumeiventris</i>	100.00%	G3G4	S3	CA
<i>Catostomus platyrhynchus</i>	23.08%	G5	S3	CA
<i>Caulanthus barnebyi</i>	100.00%	G1	S2	NV
<i>Centrocercus urophasianus</i>	2.33%	G3G4	S2S3	CA
<i>Cercyonis oetus alkalorum</i>	100.00%	G5T1	S1	NV
<i>Chaenactis carphoclinia</i> var. <i>peirsonii</i>	75.00%	G5T2	S2	CA
<i>Chaenactis douglasii</i> var. <i>alpina</i>	10.00%	G5T5	S2	CA
<i>Chaenactis parishii</i>	40.00%	G3G4	S3	CA
<i>Chaetadelpha wheeleri</i>	5.55%, 25%	G4	S2, N/A	CA, NV
<i>Chaetodipus californicus femoralis</i>	4.35%	G5T3	S3	CA
<i>Chaetodipus fallax fallax</i>	5.05%	G5T3T4	S3S4	CA
<i>Chaetodipus fallax pallidus</i>	79.31%	G5T3T4	S3S4	CA
<i>Chaetodipus penicillatus</i>	3.03%	G5	S1S2	NV
<i>Charadrius montanus</i>	59%, 100%	G3	S2S3, SNA	CA, NV
<i>Charadrius nivosus nivosus</i>	7.54%, 45%	G3T3	S2, S3B	CA, NV
<i>Charina bottae</i>	46.15%	G5	S3S4	NV
<i>Charina umbratica</i>	43.75%	G2G3	S2S3	CA
<i>Chionactis occipitalis</i>	50.00%	G5	S4	NV
<i>Chlidonias niger</i>	16.67%	G4G5	S2	CA
<i>Chloropyron tecopense</i>	100%, 100%	G2	S1, N/A	CA, NV
<i>Chorizanthe parryi</i> var. <i>parryi</i>	12.40%	G3T2	S2	CA
<i>Chorizanthe xanti</i> var. <i>leucotheca</i>	67.35%	G4T3	S3	CA

<i>Chrysothamnus greenei</i>	50.00%	G5	#N/A	NV
<i>Chylismia arenaria</i>	62.50%	G4?	S2S3	CA
<i>Chylismia claviformis</i> ssp. <i>cruciformis</i>	4.35%	G5T4	S2	CA
<i>Chylismia megalantha</i>	9.09%	G3Q	S3	NV
<i>Circus hudsonius</i>	2.65%, 18%	G5	S3, N/A	CA, NV
<i>Cirsium arizonicum</i> var. <i>tenuisectum</i>	42.86%	G5T2	S2	CA
<i>Cirsium mohavense</i>	18.75%	G3	S3	NV
<i>Cladium californicum</i>	75%, 100%	G4	S2, S2	CA, NV
<i>Clarkia xantiana</i> ssp. <i>parviflora</i>	23.53%	G4T3?	S3?	CA
<i>Claytonia megarhiza</i>	13.33%	G5	S2	CA
<i>Claytonia panamintensis</i>	100.00%	G1	S1	CA
<i>Claytonia peirsonii</i> ssp. <i>californacis</i>	100.00%	G2G3T1	S1	CA
<i>Claytonia peirsonii</i> ssp. <i>peirsonii</i>	44.44%	G2G3T2	S2	CA
<i>Claytonia peirsonii</i> ssp. <i>yorkii</i>	100.00%	G2G3T1	S1	CA
<i>Coccyzus americanus occidentalis</i>	6.85%	G5T2T3	S1	CA
<i>Colaptes chrysoides</i>	57.14%	G5	S1	CA
<i>Coleonyx switaki</i>	25.00%	G4	S1	CA
<i>Colubrina californica</i>	53.33%	G4	S2S3	CA
<i>Contopus cooperi</i>	100.00%	G4	S2B	NV
<i>Cordylanthus eremicus</i> ssp. <i>kernensis</i>	33.33%	G3T2	S2	CA
<i>Cordylanthus parviflorus</i>	38.46%	G4	S2	CA
<i>Corynorhinus townsendii</i>	27%, 26%	G4	S2, S2	CA, NV
<i>Coryphantha alversonii</i>	100.00%	G3	S3	CA
<i>Coryphantha chlorantha</i>	43.21%	G4	S3	CA
<i>Coryphantha vivipara</i> var. <i>rosea</i>	66.67%	G5T3	S1	CA
<i>Crenichthys nevadae</i>	83.33%	G1	S1	NV
<i>Crepis runcinata</i>	55.56%	G5	S3	CA
<i>Crotalus cerastes</i>	40.00%	G5	S4	NV
<i>Crotalus ruber</i>	12.85%	G4	S3	CA
<i>Crotaphytus bicinctores</i>	46.94%	G5	S4	NV
<i>Croton wigginsii</i>	90.00%	G2G3	S2	CA
<i>Cryptantha clokeyi</i>	93.75%, 100%	G3	S3, S1	CA, NV
<i>Cuniculotinus gramineus</i>	75.00%	G3G4	S3	CA
<i>Cusickiella quadricostata</i>	8.33%	G2	S2	CA
<i>Cylindropuntia munzii</i>	52.94%	G3	S1	CA
<i>Cymopterus deserticola</i>	100.00%	G2	S2	CA
<i>Cymopterus gilmanii</i>	28.57%	G3	S2	CA
<i>Cymopterus globosus</i>	100%, 46%	G3G4	S1, N/A	CA, NV
<i>Cymopterus multinervatus</i>	54.76%	G4G5	S2	CA
<i>Cymopterus nivalis</i>	100.00%	G5	S3	NV
<i>Cymopterus ripleyi</i> var. <i>saniculoides</i>	100%, 50%	G3G4T3Q	S1, S3	CA, NV

<i>Cyprinodon diabolis</i>	75.00%	G1	S1	NV
<i>Cyprinodon macularius</i>	92.65%	G1	S1	CA
<i>Cyprinodon nevadensis pectoralis</i>	100.00%	G2T1	S1	NV
<i>Cyprinodon radiosus</i>	100.00%	G1	S1	CA
<i>Cyprinodon salinus milleri</i>	100.00%	G1T1Q	S1	CA
<i>Cyprinodon salinus salinus</i>	100.00%	G1T1	S1	CA
<i>Danaus plexippus</i>	39.82%	G4	S3B	NV
<i>Dedeckera eurekaensis</i>	62.50%	G3	S3	CA
<i>Deinandra arida</i>	100.00%	G1	S1	CA
<i>Deinandra mohavensis</i>	26.56%	G2	S2	CA
<i>Delphinium inopinum</i>	5.88%	G3	S3	CA
<i>Delphinium purpusii</i>	2.86%	G3	S3	CA
<i>Delphinium recurvatum</i>	1.41%	G2?	S2?	CA
<i>Delphinium stachydeum</i>	10.34%	G5?	S3	CA
<i>Diadophis punctatus</i>	100.00%	G5	S3	NV
<i>Diadophis punctatus modestus</i>	12.00%	G5T2T3	S2?	CA
<i>Diadophis punctatus regalis</i>	66.67%	G5TNR	S2S3	CA
<i>Dieteria canescens</i> var. <i>ziegleri</i>	100.00%	G5T1	S1	CA
<i>Digitaria californica</i> var. <i>californica</i>	50.00%	G5T5	S2	CA
<i>Dinacoma caseyi</i>	100.00%	G1	S1	CA
<i>Diplacus mohavensis</i>	100.00%	G2	S2	CA
<i>Diplacus parryi</i>	50.00%	G4G5	S3	CA
<i>Diplacus pictus</i>	2.22%	G2	S2	CA
<i>Dipodomys merriami collinus</i>	11.76%	G5T2?	S1S2	CA
<i>Dipodomys panamintinus panamintinus</i>	100.00%	G5T3	S3	CA
<i>Dipodomys stephensi</i>	0.88%	G2	S2	CA
<i>Dipsosaurus dorsalis</i>	26.09%	G5	S3	NV
<i>Ditaxis claryana</i>	42.86%	G3G4	S2	CA
<i>Ditaxis serrata</i> var. <i>californica</i>	92.86%	G5T3T4	S2?	CA
<i>Draba arida</i>	66.67%	G2	S2	NV
<i>Draba asterophora</i> var. <i>asterophora</i>	33.33%	G2T2Q	S1S2	NV
<i>Draba cana</i>	100.00%	G5	S2	CA
<i>Draba lonchocarpa</i>	60.00%	G5	S2S3	CA
<i>Draba praealta</i>	80.00%	G5	S3	CA
<i>Draba saxosa</i>	80.00%	G2G3	S2S3	CA
<i>Draba sharsmithii</i>	75.00%	G2	S2	CA
<i>Draba sierrae</i>	100.00%	G3	S3	CA
<i>Draba subumbellata</i>	50.00%	G3	S1	NV
<i>Drosera anglica</i>	5.26%	G5	S2	CA
<i>Drymocallis cuneifolia</i> var. <i>cuneifolia</i>	50.00%	G2T1	S1	CA
<i>Drymocallis cuneifolia</i> var. <i>ewanii</i>	100.00%	G2T2	S2	CA

<i>Dudleya abramsii ssp. affinis</i>	69.39%	G4T2	S2	CA
<i>Dudleya densiflora</i>	1.16%	G2	S2	CA
<i>Dudleya saxosa ssp. saxosa</i>	100.00%	G4T3	S3	CA
<i>Egretta thula</i>	11.76%	G5	S4	CA
<i>Elanus leucurus</i>	0.22%	G5	S3S4	CA
<i>Elgaria coerulea palmeri</i>	20.00%	G5T4	S2S3	NV
<i>Elgaria coerulea shastensis</i>	100.00%	G5T4	S1	NV
<i>Elgaria panamintina</i>	89.47%	G3	S3	CA
<i>Elodium blandowii</i>	25.00%	G4	S2	CA
<i>Elymus salina</i>	50.00%	G4G5	S2S3	CA
<i>Empidonax traillii</i>	26.92%	G5	S1S2	CA
<i>Empidonax traillii extimus</i>	9.37%, 26%	G5T2	S1, S1B	CA, NV
<i>Emys marmorata</i>	0.71%	G3G4	S3	CA
<i>Enceliopsis covillei</i>	100.00%	G2	S2	CA
<i>Enceliopsis nudicaulis var. corrugata</i>	50%, 100%	G5T1T2Q	S1, S1	CA, NV
<i>Enneapogon desvauxii</i>	35.85%	G5	S3	CA
<i>Ensatina eschscholtzii croceater</i>	8.33%	G5T3	S3	CA
<i>Ensatina eschscholtzii klauberi</i>	13.64%	G5T2?	S3	CA
<i>Ephedra funerea</i>	100.00%	G3	S2	NV
<i>Epilobium howellii</i>	5.10%	G4	S4	CA
<i>Epilobium nevadense</i>	25.00%	G3	S2	NV
<i>Eptesicus fuscus</i>	13.00%	G5	S3S4	NV
<i>Eremarionta morongoana</i>	100.00%	G1G3	S1	CA
<i>Eremichthys acros</i>	100.00%	G1	S1	NV
<i>Eremogone ursina</i>	55.56%	G1	S1	CA
<i>Eremophila alpestris actia</i>	4.35%	G5T4Q	S4	CA
<i>Eremothera boothii ssp. boothii</i>	100.00%	G5T4	S3	CA
<i>Eremothera boothii ssp. intermedia</i>	50.00%	G5T3T4	S3	CA
<i>Eremothera nevadensis</i>	25.00%	G3	S3	NV
<i>Erethizon dorsatum</i>	7.57%	G5	S3	CA
<i>Eriastrum harwoodii</i>	90.79%	G2	S2	CA
<i>Eriastrum rosamondense</i>	100.00%	G1?	S1?	CA
<i>Eriastrum tracyi</i>	18.27%	G3Q	S3	CA
<i>Ericameria gilmanii</i>	66.67%	G2	S2	CA
<i>Erigeron compactus</i>	37.5%, 100%	G3	S3, N/A	CA, NV
<i>Erigeron miser</i>	24.14%	G3?	S3?	CA
<i>Erigeron parishii</i>	100.00%	G2	S2	CA
<i>Erigeron uncialis var. uncialis</i>	83%, 100%	G3G4T2	S2, S1	CA, NV
<i>Eriodictyon angustifolium</i>	33.33%	G5	S3	CA
<i>Eriogonum beatleyae</i>	71.43%	G3	S3	NV
<i>Eriogonum bifurcatum</i>	27.50%	G3	S3	CA

<i>Eriogonum callistum</i>	83.33%	G1	S1	CA
<i>Eriogonum contiguum</i>	64.29%	G3	S2	CA
<i>Eriogonum crosbyae</i> var. <i>crosbyae</i>	100.00%	G4T3	S3	NV
<i>Eriogonum eremicola</i>	100.00%	G1	S1	CA
<i>Eriogonum gilmanii</i>	16.67%	G3	S3	CA
<i>Eriogonum hoffmannii</i> var. <i>hoffmannii</i>	100.00%	G3T2	S2	CA
<i>Eriogonum hoffmannii</i> var. <i>robustius</i>	100.00%	G3T3	S3	CA
<i>Eriogonum intrafractum</i>	100.00%	G3	S3	CA
<i>Eriogonum kennedyi</i> var. <i>alpigenum</i>	66.67%	G4T3	S3	CA
<i>Eriogonum kennedyi</i> var. <i>austromontanum</i>	29.63%	G4T2	S2	CA
<i>Eriogonum kennedyi</i> var. <i>pinicola</i>	100.00%	G4T1	S1	CA
<i>Eriogonum mensicola</i>	100.00%	G3	S3	CA
<i>Eriogonum microthecum</i> var. <i>johnstonii</i>	50.00%	G5T2	S2	CA
<i>Eriogonum microthecum</i> var. <i>panamintense</i>	83.33%	G5T3	S3	CA
<i>Eriogonum nutans</i> var. <i>nutans</i>	17.65%	G5T3T4	S2?	CA
<i>Eriogonum ovalifolium</i> var. <i>eximium</i>	50.00%	G5T3	S2	NV
<i>Eriogonum ovalifolium</i> var. <i>vineum</i>	96.00%	G5T1	S1	CA
<i>Eriogonum ovalifolium</i> var. <i>williamsiae</i>	100.00%	G5T1	S1	NV
<i>Eriogonum robustum</i>	82.24%	G2	S2	NV
<i>Eriogonum rubricaulum</i>	25.00%	G3	S3	NV
<i>Eriogonum thornei</i>	100.00%	G1	S1	CA
<i>Eriogonum tiehmii</i>	100.00%	G1	S1	NV
<i>Eriogonum umbellatum</i> var. <i>juniporinum</i>	88.89%	G5T4	S3	CA
<i>Eriogonum umbellatum</i> var. <i>torreyanum</i>	66.67%	G5T2	S2	CA
<i>Eriogonum wrightii</i> var. <i>olanchense</i>	100.00%	G5T2	S2	CA
<i>Erioneuron pilosum</i>	80.00%	G5	S2	CA
<i>Eriophyllum mohavense</i>	100.00%	G2	S2	CA
<i>Erythranthe calcicola</i>	62.50%	G3	S3	CA
<i>Erythranthe carsonensis</i>	1.82%	G2	S2	NV
<i>Erythranthe exigua</i>	28.57%	G2	S2	CA
<i>Erythranthe purpurea</i>	40.00%	G2	S2	CA
<i>Erythranthe rhodopetra</i>	100.00%	G1	S1	CA
<i>Erythranthe shevockii</i>	30.77%	G1	S1	CA
<i>Erythranthe utahensis</i>	100.00%	G4G5	S1	CA
<i>Eschscholzia lemmonii</i> ssp. <i>kernensis</i>	1.23%	G5T2	S2	CA
<i>Eschscholzia minutiflora</i> ssp. <i>twisselmannii</i>	100.00%	G5T2	S2	CA
<i>Eucnide rupestris</i>	12.50%	G3	S1	CA
<i>Euderma maculatum</i>	32%, 57%	G4	S3, S2	CA, NV
<i>Eumops perotis californicus</i>	8.30%	G4G5T4	S3S4	CA

<i>Euparagia unidentata</i>	100.00%	G1G2	S1S2	CA
<i>Euphilotes battoides fusimaculata</i>	100.00%	G5T1	S1	NV
<i>Euphilotes bernardino inyomontana</i>	50.00%	G4T3T4	S1	NV
<i>Euphilotes enoptes aridorum</i>	20.00%	G5T1	S1	NV
<i>Euphilotes pallescens arenamontana</i>	100.00%	G3?T1	S1	NV
<i>Euphilotes pallescens calneva</i>	33.33%	G3?T1	S1	NV
<i>Euphorbia abramsiana</i>	35.65%	G4	S2	CA
<i>Euphorbia arizonica</i>	15.00%	G5	S3	CA
<i>Euphorbia exstipulata</i> var. <i>exstipulata</i>	33.33%	G5T5?	S2	CA
<i>Euphorbia jaegeri</i>	100.00%	G1	S1	CA
<i>Euphorbia misera</i>	1.54%	G5	S2	CA
<i>Euphorbia parryi</i>	100.00%	G5	S1	CA
<i>Euphydryas editha quino</i>	0.79%	G5T1T2	S1S2	CA
<i>Euphydryas editha tahoensis</i>	100.00%	G5T2T3	S1	NV
<i>Falco columbarius</i>	9.04%	G5	S3S4	CA
<i>Falco mexicanus</i>	15.00%	G5	S4	CA
<i>Falco peregrinus</i>	2.35%	G4	S3	NV
<i>Fimbristylis thermalis</i>	90%, 66%	G4	S1S2, N/A	CA, NV
<i>Fluminicola dalli</i>	100.00%	G1	S1	NV
<i>Fluminicola turbiniformis</i>	70.00%	G3	S3	NV
<i>Fluminicola virginius</i>	100.00%	G1	S1	NV
<i>Formica microphthalma</i>	100.00%	G2	S1	NV
<i>Frasera albomarginata</i> var. <i>albomarginata</i>	62.50%	G5T5	S3	CA
<i>Frasera albomarginata</i> var. <i>induta</i>	100.00%	G5T2	S1	CA
<i>Frasera pahutensis</i>	43.33%	G3Q	S3	NV
<i>Funastrum crispum</i>	100.00%	G4	S1	CA
<i>Galium angustifolium</i> ssp. <i>borregoense</i>	11.11%	G5T3?	S3?	CA
<i>Galium angustifolium</i> ssp. <i>jacinticum</i>	22.22%	G5T2?	S2?	CA
<i>Galium angustifolium</i> ssp. <i>onycense</i>	50.00%	G5T3	S3	CA
<i>Galium hilendiae</i> ssp. <i>carneum</i>	100.00%	G4T3	S3	CA
<i>Galium hilendiae</i> ssp. <i>kingstonense</i>	80.00%	G4T3	S2	CA
<i>Galium hypotrachium</i> ssp. <i>tomentellum</i>	100.00%	G5T1	S1	CA
<i>Galium proliferum</i>	53.85%	G5	S2	CA
<i>Galium wrightii</i>	100.00%	G5	S3	CA
<i>Gambelia wislizenii</i>	37.50%	G5	S4	NV
<i>Gelochelidon nilotica</i>	45.45%	G5	S1	CA
<i>Geraea viscida</i>	1.08%	G2G3	S2	CA
<i>Gila orcuttii</i>	2.00%	G2	S2	CA
<i>Gilia leptantha</i> ssp. <i>leptantha</i>	7.69%	G4T2	S2	CA
<i>Gilia ripleyi</i>	20.00%	G3	S3	NV
<i>Gilmania luteola</i>	100.00%	G2G3	S2S3	CA

<i>Glaucomys oregonensis</i>	6.90%	G5	S3	NV
<i>Glaucomys oregonensis californicus</i>	71.43%	G5T1T2	S1S2	CA
<i>Glossopetalon pungens</i>	100.00%	G2G3	S1	CA
<i>Glyceria grandis</i>	50.00%	G5	S3	CA
<i>Gonidea angulata</i>	28.57%	G3	S1	NV
<i>Gopherus agassizii</i>	65%, 23%	G3	S2S3, S2S3	CA, NV
<i>Greeneocharis circumscissa</i> var. <i>rosulata</i>	66.67%	G5T2	S2	CA
<i>Grindelia fraxinipratensis</i>	67%, 100%	G2	S1, S1	CA, NV
<i>Grusonia parishii</i>	38.46%	G3G4	S2	CA
<i>Grusonia pulchella</i>	11%, 100%	G4	S2, S3	CA, NV
<i>Gulo gulo</i>	44.44%	G4	S1	CA
<i>Gymnorhinus cyanocephalus</i>	40.00%	G3	S3	NV
<i>Hackelia sharsmithii</i>	80.00%	G3	S3	CA
<i>Haliaeetus leucocephalus</i>	2.34%, 38%	G5	S3; S2B,S4N	CA, NV
<i>Harpagonella palmeri</i>	2.04%	G4	S3	CA
<i>Helianthus deserticola</i>	100.00%	G2G3Q	S2B S4N	NV
<i>Helianthus niveus</i> ssp. <i>tephrodes</i>	33.33%	G4T2T3	S1	CA
<i>Helminthoglypta concolor</i>	33.33%	G1G2	S1S2	CA
<i>Helminthoglypta fontiphila</i>	33.33%	G1	S1	CA
<i>Helminthoglypta greggi</i>	100.00%	G1	S1	CA
<i>Helminthoglypta taylori</i>	100.00%	G1	S1	CA
<i>Heloderma suspectum cinctum</i>	33.33%	G4T4	S1	CA
<i>Hesperia miriamae longaevicola</i>	100.00%	G2G3T1T2	S3	NV
<i>Hesperia uncas fulvapalla</i>	100.00%	G4G5T1	S1	NV
<i>Hesperia uncas grandiosa</i>	37.50%	G4G5T1	S1	NV
<i>Hesperidanthus jaegeri</i>	40.00%	G2	S2	CA
<i>Heuchera hirsutissima</i>	90.48%	G3	S3	CA
<i>Heuchera parishii</i>	28.30%	G3	S3	CA
<i>Histrionicus histrionicus</i>	100.00%	G4	S1	NV
<i>Holmgrenanthe petrophila</i>	100.00%	G1	S1	CA
<i>Horkelia hispidula</i>	15.38%	G3	S3	CA
<i>Hulsea vestita</i> ssp. <i>inyoensis</i>	100.00%	G5T2T3	S1S2	CA
<i>Hulsea vestita</i> ssp. <i>pygmaea</i>	33.33%	G5T1	S1	CA
<i>Hyalella muerta</i>	100.00%	G1	S1	CA
<i>Hyalella sandra</i>	100.00%	G1	S1	CA
<i>Hydromantes platycephalus</i>	44.00%	G4	S4	CA
<i>Hydroprogne caspia</i>	33.33%	G5	S4	CA
<i>Hygrotus fontinalis</i>	100.00%	G1	S1	CA
<i>Hymenopappus filifolius</i> var. <i>eriopodus</i>	100.00%	G5T3	S2S3	CA
<i>Hymenopappus filifolius</i> var. <i>nanus</i>	75.00%	G5T4	S3	CA
<i>Icteria virens</i>	7.21%, 33%	G5	S3, SNA	CA, NV

<i>Iliamna bakeri</i>	3.13%	G4	S3	CA
<i>Imperata brevifolia</i>	36%, 75%	G3	S3, S1S2	CA, NV
<i>Ipobius robustus</i>	100.00%	G1G2	S1	CA
<i>Ipomopsis effusa</i>	100.00%	G3?	SH	CA
<i>Ipomopsis tenuifolia</i>	38.46%	G4	S2	CA
<i>Ivesia aperta</i> var. <i>aperta</i>	60.00%	G2T2	S1	NV
<i>Ivesia aperta</i> var. <i>canina</i>	100.00%	G2T1	S1	CA
<i>Ivesia argyrocoma</i> var. <i>argyrocoma</i>	37.93%	G2T2	S2	CA
<i>Ivesia arizonica</i> var. <i>arizonica</i>	100.00%	G3T3	S1	CA
<i>Ivesia callida</i>	20.00%	G1	S1	CA
<i>Ivesia campestris</i>	18.18%	G3	S3	CA
<i>Ivesia jaegeri</i>	100.00%	G2G3	S1	CA
<i>Ivesia kingii</i> var. <i>eremica</i>	100.00%	G4T1	S1	NV
<i>Ivesia kingii</i> var. <i>kingii</i>	100.00%	G4T3Q	S2	CA
<i>Ivesia patellifera</i>	50.00%	G2	S2	CA
<i>Ivesia sericoleuca</i>	50.00%	G2	S2	CA
<i>Ivesia webberi</i>	18%, 23%	G2	S1, S2	CA, NV
<i>Ixobrychus exilis</i>	17%, 20%	G4G5	S2, S2B	CA, NV
<i>Jaffuelobryum raui</i>	33.33%	G4	S2	CA
<i>Jaffuelobryum wrightii</i>	89.47%	G5	S2S3	CA
<i>Johanneshowellia crateriorum</i>	100.00%	G1	S1	NV
<i>Johanneshowellia puberula</i>	100.00%	G4?	S1	CA
<i>Juga acutifilosa</i>	60.00%	G2?	S1	NV
<i>Juncus interior</i>	100.00%	G4	S1	CA
<i>Juncus luciensis</i>	11.11%	G3	S3	CA
<i>Juncus nodosus</i>	57.14%	G5	S3	CA
<i>Juniperella mirabilis</i>	100.00%	G1	S1	CA
<i>Kobresia myosuroides</i>	33.33%	G5	S2	CA
<i>Koerberlinia spinosa</i> var. <i>tenuispina</i>	75.00%	G4T4?	S2	CA
<i>Ladeania lanceolata</i>	11.11%	G5	S2	CA
<i>Lanius ludovicianus</i>	38.53%	G4	S4	CA
<i>Larus californicus</i>	37.50%	G5	S4	CA
<i>Lasionycteris noctivagans</i>	5.13%, 19%	G3G4	S3S4, S3	CA, NV
<i>Lasiurus cinereus</i>	2.14%, 25%	G3G4	S4, S2S3	CA, NV
<i>Lasiurus frantzii</i>	7.69%	G4	S2	NV
<i>Lasiurus xanthinus</i>	22.73%	G4G5	S3	CA
<i>Laterallus jamaicensis</i>	66.67%	G3	SNR	NV
<i>Laterallus jamaicensis coturniculus</i>	6.44%	G3G4T1	S1	CA
<i>Lathyrus hitchcockianus</i>	100%, 100%	G2	S1, S2	CA, NV
<i>Layia heterotricha</i>	8.60%	G2	S2	CA
<i>Leiothlypis luciae</i>	38.46%	G5	S2S3	CA

<i>Leiothlypis virginiae</i>	14.29%	G5	S2	CA
<i>Lemmiscus curtatus</i>	22.22%	G5	S3	NV
<i>Lepidium integrifolium</i>	100.00%	G2G3T2T3	S1	NV
<i>Lepidium nanum</i>	16.67%	G3	S3	NV
<i>Lepus americanus tahoensis</i>	40%, 29%	G5T3T4Q	S2, S3	CA, NV
<i>Lepus townsendii townsendii</i>	100.00%	G5T5	S3?	CA
<i>Lewisia brachycalyx</i>	16.67%	G4	S2	CA
<i>Lewisia longipetala</i>	28.57%	G2	S2	CA
<i>Lilium parryi</i>	29.55%	G3	S3	CA
<i>Limenitis archippus lahontani</i>	37.50%	G5T1T2	S1S2	NV
<i>Linanthus bernardinus</i>	100.00%	G1	S1	CA
<i>Linanthus concinnus</i>	52.17%	G2	S2	CA
<i>Linanthus jaegeri</i>	66.67%	G2	S2	CA
<i>Linanthus killipii</i>	53.85%	G1	S1	CA
<i>Linanthus maculatus ssp. emaculatus</i>	100.00%	G2T1	S1	CA
<i>Linanthus maculatus ssp. maculatus</i>	100.00%	G2T2	S2	CA
<i>Linum puberulum</i>	25.00%	G5	S2	CA
<i>Lithobates onca</i>	5.56%	G1G2	S1	NV
<i>Lithobates pipiens</i>	100%, 8.33%	G5	S2, S2S3	CA, NV
<i>Loeflingia squarrosa var. artemisiarum</i>	68.75%	G5T3	S2	CA
<i>Lomatium grayi</i>	25.00%	G5	S1S2	CA
<i>Lomatium shevockii</i>	100.00%	G2	S2	CA
<i>Lupinus albifrons var. medius</i>	40.00%	G4T2	S2	CA
<i>Lupinus duranii</i>	100.00%	G2	S2	CA
<i>Lupinus gracilentus</i>	6.25%	G3	S3	CA
<i>Lupinus holmgrenianus</i>	100%, 100%	G2	S2, S2	CA, NV
<i>Lupinus magnificus var. hesperius</i>	100.00%	G3T1Q	S1	CA
<i>Lupinus magnificus var. magnificus</i>	100.00%	G3T3	S3	CA
<i>Lupinus padre-crowleyi</i>	100.00%	G2	S2	CA
<i>Lupinus peirsonii</i>	75.00%	G3	S3	CA
<i>Lupinus pusillus var. intermontanus</i>	10.00%	G5T5?	S2	CA
<i>Lycium parishii</i>	73.33%	G4	S1	CA
<i>Macrobaenetes valgum</i>	100.00%	G1G2	S1S2	CA
<i>Macrotus californicus</i>	58.14%	G3G4	S3	CA
<i>Malperia tenuis</i>	72.41%	G4?	S2?	CA
<i>Margaritifera falcata</i>	2.86%, 7.14%	G4G5	S1S2, S1	CA, NV
<i>Marina orcuttii var. orcuttii</i>	100.00%	G2G3T1T2	S2?	CA
<i>Martes caurina</i>	12.50%	G4G5	S2S3	NV
<i>Martes caurina sierrae</i>	14.46%	G4G5T3	S3	CA
<i>Matelea parvifolia</i>	33.33%	G5	S3	CA
<i>Maurandella antirrhiniflora</i>	100.00%	G5	S2	CA

<i>Meesia triquetra</i>	21.43%	G5	S1	NV
<i>Meesia uliginosa</i>	38.00%	G5	S3	CA
<i>Melanerpes uropygialis</i>	29.17%	G5	S1	CA
<i>Melospiza melodia</i>	1.07%, 28%	G5	S3?, N/A	CA, NV
<i>Melozone crissalis eremophilus</i>	100.00%	G4G5T2	S2	CA
<i>Menodora scabra</i> var. <i>scabra</i>	46.15%	G5T4T5	S3	CA
<i>Menodora spinescens</i> var. <i>mohavensis</i>	100.00%	G4T2	S2	CA
<i>Mentzelia hirsutissima</i>	22.86%	G4?	S3	CA
<i>Mentzelia inyoensis</i>	67%, 100%	G2	S1, S1	CA, NV
<i>Mentzelia leucophylla</i>	100.00%	G1	S1	NV
<i>Mentzelia mollis</i>	100.00%	G2	S1	NV
<i>Mentzelia polita</i>	37.50%	G2G3	S2?	CA
<i>Mentzelia pterosperma</i>	33%, 100%	G4	S1S2, N/A	CA, NV
<i>Mentzelia puberula</i>	50.00%	G5	S2	CA
<i>Mentzelia torreyi</i>	93.33%	G4	S2	CA
<i>Mentzelia tridentata</i>	97.06%	G3	S3	CA
<i>Mertensia oblongifolia</i> var. <i>oblongifolia</i>	14.29%	G5T5	S3	CA
<i>Micrathene whitneyi</i>	16.67%	G5	S1	CA
<i>Microdipodops pallidus</i>	62.86%	G3	S2	NV
<i>Micromonolepis pusilla</i>	50.00%	G5	S3?	CA
<i>Microtus californicus mohavensis</i>	100.00%	G5T1	S1	CA
<i>Microtus californicus vallicola</i>	100.00%	G5T3	S3	CA
<i>Miloderes amargosensis</i>	100.00%	G1	S1	NV
<i>Mirabilis coccinea</i>	20.00%	G4G5	S2	CA
<i>Monardella beneolens</i>	100.00%	G2	S2	CA
<i>Monardella boydii</i>	100.00%	G1?Q	S1?	CA
<i>Monardella eremicola</i>	80.00%	G3Q	S3	CA
<i>Monardella linoides</i> ssp. <i>oblonga</i>	35.42%	G5T2	S2	CA
<i>Monardella nana</i> ssp. <i>leptosiphon</i>	11.11%	G4G5T2Q	S2	CA
<i>Monardella robisonii</i>	100.00%	G3	S3	CA
<i>Muhlenbergia appressa</i>	60.87%	G4	S3	CA
<i>Muhlenbergia arsenei</i>	66.67%	G4	S2?	CA
<i>Muhlenbergia pauciflora</i>	75.00%	G5	S2	CA
<i>Muhlenbergia utilis</i>	62.50%	G4	S2S3	CA
<i>Munroa squarrosa</i>	75%, 100%	G5	S2, N/A	CA, NV
<i>Myiarchus tyrannulus</i>	50.00%	G5	S3	CA
<i>Myotis californicus</i>	22.00%	G5	S3S4	NV
<i>Myotis ciliolabrum</i>	16%, 27%	G5	S3, S3S4	CA, NV
<i>Myotis evotis</i>	4.10%, 18%	G5	S3, S3	CA, NV
<i>Myotis lucifugus</i>	18.75%	G3G4	S2S3	NV
<i>Myotis thysanodes</i>	7.05%, 16%	G4	S3, S2	CA, NV

<i>Myotis velifer</i>	14.29%	G4G5	S1	CA
<i>Myotis volans</i>	10%, 22%	G4G5	S3, S3S4	CA, NV
<i>Myotis yumanensis</i>	3.44%	G5	S4	CA
<i>Myurella julacea</i>	50.00%	G5	S2	CA
<i>Nama demissa var. covillei</i>	100.00%	G5T3	S3	CA
<i>Nama dichotoma var. dichotoma</i>	100.00%	G5T5?	S1	CA
<i>Nardia hiroshii</i>	100.00%	G4G5	S1	CA
<i>Navarretia fossalis</i>	1.67%	G2	S2	CA
<i>Navarretia peninsularis</i>	19.23%	G3	S2	CA
<i>Navarretia setiloba</i>	1.96%	G2	S2	CA
<i>Nemacaulis denudata var. gracilis</i>	90.91%	G3G4T3?	S2	CA
<i>Nemacladus inyoensis</i>	70.59%	G3	S3	CA
<i>Neotamias amoenus celeris</i>	66.67%	G5T1	S2	NV
<i>Neotoma albigula venusta</i>	50.00%	G5T3T4	S1S2	CA
<i>Neotoma lepida intermedia</i>	35.94%	G5T3T4	S3S4	CA
<i>Nitrophila mohavensis</i>	100%, 100%	G1	S1, S1	CA, NV
<i>Notiosorex crawfordi</i>	21.74%	G4	S3	NV
<i>Numenius americanus</i>	50.00%	G5	S2S3B	NV
<i>Nycticorax nycticorax</i>	6.90%	G5	S4	CA
<i>Nyctinomops femorosaccus</i>	8.57%	G5	S3	CA
<i>Nyctinomops macrotis</i>	8.33%	G5	S3	CA
<i>Ochlodes yuma lutea</i>	50.00%	G4TNR	S2	NV
<i>Ochotona princeps</i>	24.84%	G5	S2	NV
<i>Ochotona princeps schisticeps</i>	54.15%	G5T4	S2S4	CA
<i>Oenothera longissima</i>	100.00%	G4	S1	CA
<i>Oliarces clara</i>	100.00%	G1G3	S2	CA
<i>Oncorhynchus clarkii henshawi</i>	11%, 28%	G5T3	S1, S3	CA, NV
<i>Oncorhynchus clarkii utah</i>	6.67%	G5T4	S1	NV
<i>Onychomys torridus tularensis</i>	8.33%	G5T1T2	S1S2	CA
<i>Opuntia basilaris var. brachyclada</i>	50.28%	G5T3	S3	CA
<i>Oreocarya roosiorum</i>	83.33%	G2	S2	CA
<i>Oreocarya schoolcraftii</i>	87.50%	G3	S3	NV
<i>Oreohelix hemphilli</i>	23.53%	G2	S2	NV
<i>Oreonana vestita</i>	43.40%	G3	S3	CA
<i>Orobanche ludoviciana var. arenosa</i>	100.00%	G5T5	S2	CA
<i>Orobanche valida ssp. valida</i>	50.00%	G4T2	S2	CA
<i>Oryctes nevadensis</i>	96%, 50%	G3	S2, S2S3	CA, NV
<i>Ovis canadensis nelsoni</i>	80%, 4%	G4T4	S3, S4	CA, NV
<i>Ovis canadensis nelsoni pop. 2</i>	50.00%	G4T3Q	S2	CA
<i>Ovis canadensis sierrae</i>	100.00%	G4T2	S2	CA
<i>Oxytheca watsonii</i>	75%, 100%	G3?	S1, S3?	CA, NV

<i>Oxytropis oreophila</i> var. <i>juniperina</i>	100.00%	G5T4T5	S1	CA
<i>Oxytropis oreophila</i> var. <i>oreophila</i>	50.00%	G5T4T5	S2	CA
<i>Packera bernardina</i>	43.48%	G2	S2	CA
<i>Palaeoxenus dohrni</i>	50.00%	G3?	S3?	CA
<i>Palafoxia arida</i> var. <i>gigantea</i>	100.00%	G5T3?	S2	CA
<i>Pandion haliaetus</i>	2.33%, 22%	G5	S4, S4	CA, NV
<i>Panicum hirticaule</i> ssp. <i>hirticaule</i>	66.67%	G5T5	S2	CA
<i>Parastrellus hesperus</i>	26.23%	G5	S3S4	NV
<i>Parnassia parviflora</i>	100.00%	G5?	S2	CA
<i>Paruroctonus arenicola</i> <i>arenicola</i>	100.00%	GNRTNR	S1	NV
<i>Pedicularis crenulata</i>	100.00%	G4	S1	CA
<i>Pediomelum castoreum</i>	100.00%	G3	S2	CA
<i>Pelecanus erythrorhynchos</i>	4.88%, 31%	G4	S1S2, S2B	CA, NV
<i>Pelecanus occidentalis californicus</i>	3.45%	G4T3T4	S3	CA
<i>Pellaea truncata</i>	50.00%	G5	S3	CA
<i>Pelocoris biimpressus</i>	100.00%	G1G3	S1S2	CA
<i>Penstemon albomarginatus</i>	100%, 69%	G2	S1, S2	CA, NV
<i>Penstemon calcareus</i>	72.73%	G3?	S3?	CA
<i>Penstemon californicus</i>	80.00%	G3	S2	CA
<i>Penstemon floribundus</i>	85.71%	G2	S2	NV
<i>Penstemon fruticiformis</i> ssp. <i>amargosae</i>	26.32%	G4T3	S2	NV
<i>Penstemon fruticiformis</i> var. <i>amargosae</i>	100.00%	G4T3	S2	CA
<i>Penstemon janishiae</i>	100.00%	G4	#N/A	NV
<i>Penstemon pahutensis</i>	100%, 86%	G3	S1, S3	CA, NV
<i>Penstemon palmeri</i> var. <i>macranthus</i>	9.09%	G4G5T2?	S2?	NV
<i>Penstemon pseudospectabilis</i> ssp. <i>pseudospectabilis</i>	30.00%	G4G5T4	S3	CA
<i>Penstemon pudicus</i>	50.00%	G1G2	S1	NV
<i>Penstemon stephensii</i>	81.25%	G3?	S3?	CA
<i>Penstemon sudans</i>	2.13%	G4	S4	CA
<i>Penstemon thompsoniae</i>	20.00%	G4	S1	CA
<i>Penstemon utahensis</i>	50.00%	G4	S2	CA
<i>Perdita frontalis</i>	100.00%	G1G2	S1S2	CA
<i>Perideridia parishii</i> ssp. <i>parishii</i>	55.17%	G4T3T4	S2	CA
<i>Perityle inyoensis</i>	68.75%	G2	S2	CA
<i>Perityle villosa</i>	100.00%	G2	S2	CA
<i>Perognathus alticola inexpectatus</i>	100.00%	G2T1T2	S1S2	CA
<i>Perognathus inornatus</i>	1.09%	G2G3	S2S3	CA
<i>Perognathus longimembris bangsi</i>	90.00%	G5T2	S2	CA
<i>Perognathus longimembris brevinasus</i>	36.36%	G5T2	S1S2	CA
<i>Perognathus mollipilosus xanthonotus</i>	100.00%	G5T2	S1S2	CA
<i>Petalonyx linearis</i>	21.74%	G4	S3?	CA

<i>Petalonyx thurberi</i> ssp. <i>gilmanii</i>	100.00%	G5T2	S2	CA
<i>Petrophytum caespitosum</i> ssp. <i>acuminatum</i>	33.33%	G5T2	S2	CA
<i>Phacelia anelsonii</i>	16.67%	G3	S2	CA
<i>Phacelia barnebyana</i>	100.00%	G3?	S2	CA
<i>Phacelia beatleyae</i>	5.26%	G3	S2	NV
<i>Phacelia coerulea</i>	33.33%	G5	S2	CA
<i>Phacelia filiae</i>	58.33%	G3	S3	NV
<i>Phacelia glaberrima</i>	100.00%	G3?	S3?	NV
<i>Phacelia gymnoclada</i>	45.45%	G4	#N/A	NV
<i>Phacelia inconspicua</i>	25.00%	G2	S1	NV
<i>Phacelia inundata</i>	14.29%	G3	S2?	NV
<i>Phacelia inyoensis</i>	100.00%	G2	S2	CA
<i>Phacelia monoensis</i>	11.11%	G3	S2	CA
<i>Phacelia mustelina</i>	100.00%	G3	S2	CA
<i>Phacelia nashiana</i>	92.16%	G3	S3	CA
<i>Phacelia novenmillensis</i>	40.91%	G3	S3	CA
<i>Phacelia parishii</i>	75.00%	G2G3	S1	CA
<i>Phacelia perityloides</i> var. <i>jaegeri</i>	100.00%	G4T2	S2	CA
<i>Phainopepla nitens</i>	0.45%	G5	S3	NV
<i>Phlox dolichantha</i>	37.14%	G2	S2	CA
<i>Pholisma sonora</i>	100.00%	G2	S2	CA
<i>Pholistoma auritum</i> var. <i>arizonicum</i>	36.36%	G5T4?	S3	CA
<i>Phrynosoma blainvillii</i>	8.71%	G3G4	S3S4	CA
<i>Phrynosoma mcallii</i>	59.72%	G3	S2	CA
<i>Phrynosoma platyrhinos</i>	38.30%	G5	S4	NV
<i>Phyciodes pulchella shoshoni</i>	100.00%	G5T2	S2	NV
<i>Phyciodes pulchella vallis</i>	100.00%	G5T4	S2	NV
<i>Physalis lobata</i>	41.67%	G5	S1S2	CA
<i>Physaria chambersii</i>	75.00%	G5	S3	CA
<i>Physaria kingii</i> ssp. <i>bernardina</i>	33.33%	G5T1	S1	CA
<i>Physaria ludoviciana</i>	50.00%	G5	S1	CA
<i>Physocarpus alternans</i>	70.00%	G4	S3	CA
<i>Picoides arcticus</i>	10.53%	G5	S2	CA
<i>Pinus albicaulis</i>	10.53%	G3G4	S3	NV
<i>Pinus ponderosa</i> ssp. <i>washoensis</i>	66.67%	G3Q	S2	NV
<i>Piptatherum shoshoneanum</i>	100.00%	G2G3	S1	NV
<i>Piranga rubra</i>	42.00%	G5	S1	CA
<i>Plagiobothrys glomeratus</i>	66.67%	G2G3	S2	NV
<i>Plagiobothrys parishii</i>	100.00%	G1	S1	CA
<i>Plebulina emigdionis</i>	37.50%	G1G2	S1S2	CA
<i>Plegadis chihi</i>	4.89%, 41%	G5	S3S4, S3B	CA, NV

<i>Plestiodon gilberti rubricaudatus</i>	65.91%	G5T4	S2S3	NV
<i>Poa atropurpurea</i>	5.88%	G2	S2	CA
<i>Poa lettermanii</i>	100.00%	G4	S3	CA
<i>Pohlia tundrae</i>	25.00%	G3	S3	CA
<i>Polemonium chartaceum</i>	43%, 100%	G2	S2, S1	CA, NV
<i>Polioptila californica californica</i>	0.32%	G4G5T3Q	S2	CA
<i>Polioptila melanura</i>	58.33%	G5	S3S4	CA
<i>Polites sabuleti genoa</i>	33.33%	G5T3T4	S1	NV
<i>Polyctenium fremontii var. bisulcatum</i>	100.00%	G4TH	SNA	NV
<i>Polyctenium williamsiae</i>	40.00%	G2Q	S1	CA
<i>Polygala acanthoclada</i>	8.70%	G4	S2S3	CA
<i>Polygala intermontana</i>	100.00%	G4	S2	CA
<i>Polygala subspinosa</i>	10.34%	G4?	S3	CA
<i>Populus angustifolia</i>	100.00%	G5	S2	CA
<i>Potamogeton robbinsii</i>	71.43%	G5	S3	CA
<i>Potentilla morefieldii</i>	40.00%	G2	S2	CA
<i>Potentilla rimicola</i>	60.00%	G2	S1	CA
<i>Prosopium williamsoni</i>	20%, 100%	G5	S3, S3	CA, NV
<i>Prunus eremophila</i>	75.00%	G2	S2	CA
<i>Pseudocopaeodes eunus flavus</i>	60.00%	G3T3	S1	NV
<i>Pseudocopaeodes eunus obscurus</i>	72.73%	G3T1	S1	NV
<i>Pseudocotalpa andrewsi</i>	100.00%	G1	S1	CA
<i>Pseudocotalpa giulianii</i>	100.00%	G1	S1	NV
<i>Pseudorontium cyathiferum</i>	22.22%	G4G5	S1	CA
<i>Psiloscops flammeolus</i>	12%, 33%	G4	S2S4, S3	CA, NV
<i>Puccinellia parishii</i>	100.00%	G3	S1	CA
<i>Puccinellia simplex</i>	17.14%	G3	S2	CA
<i>Pyrgulopsis aloba</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis aurata</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis basiglans</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis bifurcata</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis bruesi</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis bryantwalkerii</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis crystalis</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis dixensis</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis eremica</i>	36.36%	G2	S2	CA
<i>Pyrgulopsis erythropoma</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis fairbanksensis</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis gibba</i>	52.78%	G3	S3	NV
<i>Pyrgulopsis isolatus</i>	100.00%	G1	SX	NV
<i>Pyrgulopsis kolobensis</i>	2.94%	G5	S3	NV

<i>Pyrgulopsis licina</i>	100.00%	GNR	S1	NV
<i>Pyrgulopsis limaria</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis lockensis</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis longiglans</i>	93.33%	G2	S2	NV
<i>Pyrgulopsis micrococcus</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis militaris</i>	50.00%	G1	S1	NV
<i>Pyrgulopsis nanus</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis owensensis</i>	100.00%	G1G2	S1S2	CA
<i>Pyrgulopsis papillata</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis pellita</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis perturbata</i>	100.00%	G1	S1	CA
<i>Pyrgulopsis pisteri</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis ruinosa</i>	100.00%	GX	S1	NV
<i>Pyrgulopsis sadai</i>	50.00%	G2	S1S2	NV
<i>Pyrgulopsis sanchezi</i>	100.00%	GNR	S2	NV
<i>Pyrgulopsis turbatrix</i>	14.29%	G3	S1	NV
<i>Pyrgulopsis umbilicata</i>	100.00%	G1Q	S1	NV
<i>Pyrgulopsis varneri</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis villacampae</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis wongi</i>	81%, 50%	G2	S2, S1	CA, NV
<i>Pyrocephalus rubinus</i>	32.80%	G5	S2S3	CA
<i>Pyrocoma uniflora</i> var. <i>gossypina</i>	14.29%	G5T1	S1	CA
<i>Rallus obsoletus yumanensis</i>	41%, 62%	G3T3	S1S2, S1B	CA, NV
<i>Rana draytonii</i>	0.12%	G2G3	S2S3	CA
<i>Rana luteiventris</i> pop. 3	0.66%	G4T2T4Q	S2S3	NV
<i>Rana muscosa</i>	18.03%	G1	S1	CA
<i>Rana sierrae</i>	6.69%	G1	S1	CA
<i>Ranunculus hydrocharoides</i>	100.00%	G4	S1	CA
<i>Rhamnus alnifolia</i>	52.94%	G5	S3	CA
<i>Rhinichthys osculus nevadensis</i>	100.00%	G5T1	S1	NV
<i>Rhinichthys osculus</i> ssp. 12	100.00%	G5T1	S1	CA
<i>Rhinichthys osculus</i> ssp. 2	100.00%	G5T1T2Q	S1S2	CA
<i>Rhinichthys osculus</i> ssp. 5	100.00%	G5T1	S1	NV
<i>Rhinichthys osculus</i> ssp. 6	100.00%	G5T1	S1	NV
<i>Riparia riparia</i>	2.46%, 67%	G5	S2, S2B	CA, NV
<i>Rynchops niger</i>	1.34%	G5	S2	CA
<i>Sabulina stricta</i>	75.00%	G5	S3	CA
<i>Salix brachycarpa</i> var. <i>brachycarpa</i>	100.00%	G5T5	S2	CA
<i>Salix nivalis</i>	70.00%	G5	S2	CA
<i>Saltugilia latimeri</i>	93.33%	G3	S3	CA
<i>Salvadora hexalepis virgultea</i>	2.50%	G5T4	S2S3	CA

<i>Salvia greatae</i>	100.00%	G2G3	S2S3	CA
<i>Sanvitalia abertii</i>	44.07%	G5	S2S3	CA
<i>Sarcobatus baileyi</i>	50%, 50%	G4	S1, N/A	CA, NV
<i>Satyrrium sylvinus megapallidum</i>	33.33%	G3G4T2T4	S2	NV
<i>Sauromalus ater</i>	75.44%	G5	S3	NV
<i>Scaphiopus couchii</i>	83.33%	G5	S2	CA
<i>Sceloporus graciosus graciosus</i>	100.00%	G5T5	S3	CA
<i>Schoenus nigricans</i>	57%, 67%	G4	S2, N/A	CA, NV
<i>Sclerocactus blainei</i>	50.00%	G1G2Q	S1	NV
<i>Sclerocactus johnsonii</i>	100.00%	G3	S2	CA
<i>Sclerocactus polycanistrus</i>	62.50%	G3	S2S3	NV
<i>Scutellaria bolanderi ssp. austromontana</i>	3.57%	G4T3	S3	CA
<i>Scutellaria galericulata</i>	3.13%	G5	S2	CA
<i>Selaginella eremophila</i>	18.99%	G4	S2S3	CA
<i>Senna covesii</i>	15.15%	G5	S3	CA
<i>Setophaga petechia</i>	7.86%, 16%	G5	S3S4, N/A	CA, NV
<i>Sidalcea covillei</i>	100.00%	G2	S2	CA
<i>Sidalcea hickmanii ssp. parishii</i>	10.53%	G3T1	S1	CA
<i>Sidalcea neomexicana</i>	16.67%	G4	S2	CA
<i>Sidotheca emarginata</i>	39.29%	G3	S3	CA
<i>Sigmodon hispidus eremicus</i>	83.33%	G5T2T3	S2	CA
<i>Silene krantzii</i>	33.33%	G1	S1	CA
<i>Silene nachlingerae</i>	8.33%	G2	S2	NV
<i>Silene oregana</i>	8.00%	G4	S2	CA
<i>Siphateles bicolor euchila</i>	100.00%	G4T1Q	S1	NV
<i>Siphateles bicolor mohavensis</i>	83.33%	G4T1	S1	CA
<i>Siphateles bicolor newarkensis</i>	100.00%	G4T1	S1	NV
<i>Siphateles bicolor snyderi</i>	100.00%	G4T1	S1	CA
<i>Siphateles bicolor ssp. 4</i>	60.00%	G4T1Q	S1	NV
<i>Siphateles bicolor ssp. 6</i>	100.00%	G4T1	S1	NV
<i>Siphateles bicolor ssp. 7</i>	100.00%	G4T1Q	S1	NV
<i>Siphateles bicolor ssp. 9</i>	100.00%	G4T1Q	S1	NV
<i>Sisyrinchium funereum</i>	100%, 100%	G2	S2, S1S2	CA, NV
<i>Solorina spongiosa</i>	100.00%	G4G5	S1	CA
<i>Sorex merriami</i>	20.00%	G4	S3	NV
<i>Sorex navigator</i>	10.34%	G5	S2	NV
<i>Sorex tenellus</i>	7.69%	G4	S2	NV
<i>Sorex trowbridgii</i>	100.00%	G5	S2	NV
<i>Spea hammondii</i>	0.32%	G2G3	S3	CA
<i>Spermolepis gigantea</i>	100.00%	G2G3	SH	CA
<i>Sphaeralcea rusbyi var. eremicola</i>	44.44%	G4T2	S2	CA

<i>Sphaeromeria argentea</i>	100.00%	G3G4	S1?	NV
<i>Sphaeromeria potentilloides</i> var. <i>nitrophila</i>	100.00%	G5T4?	S2	CA
<i>Sphenopholis obtusata</i>	25.00%	G5	S2	CA
<i>Spinus lawrencei</i>	7.43%	G3G4	S4	CA
<i>Spiranthes infernalis</i>	100.00%	G2	S1	NV
<i>Spizella atrogularis</i>	11.11%	G5	S2B	NV
<i>Spizella breweri</i>	100%, 14%	G5	S4, S3B	CA, NV
<i>Stenelmis calida calida</i>	100.00%	GNRT1	S1	NV
<i>Stenopelmatus cahuilensis</i>	100.00%	G1G2	S1S2	CA
<i>Stipa arida</i>	38.10%	G5	S3?	CA
<i>Streptanthus bernardinus</i>	33.33%	G3G4	S3S4	CA
<i>Streptanthus campestris</i>	16.98%	G3	S3	CA
<i>Streptanthus cordatus</i> var. <i>piutensis</i>	25.00%	G5T1	S1	CA
<i>Streptanthus medeirosii</i>	33.33%	G1	S1	CA
<i>Streptanthus oliganthus</i>	25.00%	G3	S3	CA
<i>Strix occidentalis occidentalis</i>	22.22%	G3G4T2T3	S1	NV
<i>Stygobromus myersae</i>	100.00%	G1G2	S1S2	CA
<i>Stygobromus sheldoni</i>	100.00%	G1	S1	CA
<i>Stygobromus sierrensis</i>	100.00%	G1	S1	CA
<i>Suaeda occidentalis</i>	33.33%	G5	S2	CA
<i>Symphyotrichum defoliatum</i>	3.45%	G2	S2	CA
<i>Symphyotrichum greatae</i>	14.71%	G2	S2	CA
<i>Tadarida brasiliensis</i>	17.86%	G5	S4	NV
<i>Tamiasciurus douglasii</i>	4.76%	G5	S5	NV
<i>Taraxacum californicum</i>	18.18%	G1G2	S1S2	CA
<i>Taxidea taxus</i>	6.64%, 14%	G5	S3, N/A	CA, NV
<i>Tetracoccus ilicifolius</i>	100.00%	G2	S2	CA
<i>Tetradymia tetrameres</i>	80%, 20%	G4	S2, N/A	CA, NV
<i>Teucrium cubense</i> ssp. <i>depressum</i>	100.00%	G4G5T3T4	S2	CA
<i>Thamnophis hammondii</i>	9.09%	G4	S3S4	CA
<i>Thelypodium integrifolium</i> ssp. <i>complanatum</i>	100.00%	G5T4T5	S2	CA
<i>Thomomys bottae abstrusus</i>	100.00%	G5T1	S1	NV
<i>Thysanocarpus rigidus</i>	50.00%	G1G2	S1	CA
<i>Tidestromia eliassoniana</i>	100.00%	G5	S2	CA
<i>Tonestus alpinus</i>	100.00%	G2	S2	NV
<i>Tonestus graniticus</i>	100.00%	G1	S1	NV
<i>Tortella alpicola</i>	50.00%	G5?	S1	CA
<i>Townsendia condensata</i>	30.00%	G4	S3	CA
<i>Townsendia leptotes</i>	37.50%	G4	S2	CA
<i>Toxostoma bendirei</i>	61.54%	G4	S3	CA

<i>Toxostoma crissale</i>	20.00%	G5	S3	CA
<i>Toxostoma lecontei</i>	67%, 6.67%	G4	S3, S2	CA, NV
<i>Trichinorhipis knullii</i>	100.00%	G1	S1	CA
<i>Trichophorum pumilum</i>	33.33%	G5	S3	CA
<i>Trichostema austromontanum ssp. compactum</i>	100.00%	G3G4T1	S1	CA
<i>Trifolium dedeckerae</i>	78.57%	G2	S2	CA
<i>Trifolium rollinsii</i>	36.36%	G2G3Q	S2	NV
<i>Triglochin palustris</i>	62.50%	G5	S2	CA
<i>Tripterocalyx crux-maltae</i>	50.00%	G4?	#N/A	NV
<i>Triteleia piutensis</i>	33.33%	G1	S1	CA
<i>Tryonia angulata</i>	100.00%	G1	S1	NV
<i>Tryonia elata</i>	100.00%	G1	S1	NV
<i>Tryonia ericae</i>	100.00%	G1	S1	NV
<i>Tryonia margae</i>	100.00%	G1	S1	CA
<i>Tryonia monitorae</i>	100.00%	G1	S1	NV
<i>Tryonia rowlandsi</i>	100.00%	G1	S1	CA
<i>Tryonia variegata</i>	100.00%	G2	S2	NV
<i>Uma inornata</i>	100.00%	G1Q	S1	CA
<i>Uma notata</i>	55.56%	G3	S2	CA
<i>Uma scoparia</i>	88.24%	G3G4	S3S4	CA
<i>Viola lithion</i>	40.00%	G1G2	S1	NV
<i>Viola pinetorum ssp. grisea</i>	17.86%	G4G5T3	S3	CA
<i>Viola purpurea ssp. aurea</i>	27.27%	G5T2	S2	CA
<i>Vireo bellii pusillus</i>	3.76%	G5T2	S2	CA
<i>Vulpes macrotis</i>	100.00%	G4	S3	NV
<i>Vulpes vulpes necator</i>	4.55%	G5T1T2	S1	CA
<i>Wislizenia refracta ssp. palmeri</i>	100.00%	G5T3T5	S1	CA
<i>Wislizenia refracta ssp. refracta</i>	100.00%	G5T5?	S1	CA
<i>Woodsia plummerae</i>	50.00%	G5	S2	CA
<i>Xanthocephalus xanthocephalus</i>	23%, 27%	G5	S3, N/A	CA, NV
<i>Xantusia gracilis</i>	50.00%	G1	S1	CA
<i>Xerospermophilus mohavensis</i>	99.17%	G2G3	S2S3	CA
<i>Xerospermophilus tereticaudus chlorus</i>	100.00%	G5T2Q	S2	CA
<i>Xylorhiza cognata</i>	100.00%	G2	S2	CA
<i>Xylorhiza orcuttii</i>	10.71%	G3?	S2	CA
<i>Xyrauchen texanus</i>	12.50%	G1	S1S2	CA
<i>Zapus princeps</i>	12.31%	G5	S2	NV
<i>Zeltnera namophila</i>	100.00%	G2Q	S1	NV

Appendix B. List of species recorded on proposed lithium projects in California and Nevada with the percent of their records found within the proposed projects, their G rank and S rank. Four species were

recorded in both states, the first value in the cell corresponds to data from California and the second value from Nevada.

Species	Inside project	G Rank	S Rank	State
<i>Accipiter cooperii</i>	1.14%	G5	S4	CA
<i>Agelaius tricolor</i>	0.54%	G1G2	S1S2	CA
<i>Anaxyrus californicus</i>	1.59%	G2G3	S2S3	CA
<i>Anaxyrus nevadensis</i>	100.00%	G1	S1	NV
<i>Anaxyrus sp. 2</i>	100.00%	GU	S2	NV
<i>Androstephium breviflorum</i>	0.87%	G4	S2?	CA
<i>Antigone canadensis tabida</i>	3.57%	G5T5	S2B, S2M	NV
<i>Antrozous pallidus</i>	0.82%, 4.95%	G4	S3, S3	CA, NV
<i>Aquila chrysaetos</i>	0.34%	G5	S3	CA
<i>Arctostaphylos rainbowensis</i>	4.94%	G2	S2	CA
<i>Asclepias eastwoodiana</i>	9.09%	G2	S2S3	NV
<i>Aspidoscelis hyperythra</i>	0.39%	G5	S2S3	CA
<i>Athene cunicularia</i>	0.63%	G4	S3	CA
<i>Boechera shockleyi</i>	5.88%	G3	S3	NV
<i>Brachylagus idahoensis</i>	2.51%	G4	S3	NV
<i>Branchinecta lynchi</i>	0.40%	G3	S3	CA
<i>Branchinecta mesovallensis</i>	1.52%	G2	S2S3	CA
<i>Brodiaea orcuttii</i>	1.01%	G2	S2	CA
<i>Buteo swainsoni</i>	0.24%	G5	S3	CA
<i>Castela emoryi</i>	2.63%	G3G4	S2S3	CA
<i>Charadrius montanus</i>	5.13%	G3	S2S3	CA
<i>Charadrius nivosus nivosus</i>	2.22%, 9.09%	G3T3	S2, S3B	CA, NV
<i>Chorizanthe polygonoides var. longispina</i>	2.65%	G5T3	S3	CA
<i>Circus hudsonius</i>	0.42%	G5	S3	CA
<i>Cirsium mohavense</i>	6.25%	G3	S3	NV
<i>Corynorhinus townsendii</i>	1.61%, 4.41%	G4	S2, S2	CA, NV
<i>Crenichthys nevadae</i>	16.67%	G1	S1	NV
<i>Cyprinodon macularius</i>	10.77%	G1	S1	CA
<i>Deinandra mohavensis</i>	1.59%	G2	S2	CA
<i>Downingia pusilla</i>	2.17%	GU	S2	CA
<i>Elanus leucurus</i>	0.24%	G5	S3S4	CA
<i>Empidonax traillii extimus</i>	3.17%	G5T2	S1	CA
<i>Emys marmorata</i>	0.09%	G3G4	S3	CA
<i>Enceliopsis covillei</i>	37.50%	G2	S2	CA
<i>Eptesicus fuscus</i>	1.00%	G5	S3S4	NV
<i>Eriogonum hoffmannii var. hoffmannii</i>	33.33%	G3T2	S2	CA

<i>Eriogonum tiehmii</i>	100.00%	G1	S1	NV
<i>Euderma maculatum</i>	14.29%	G4	S2	NV
<i>Eumops perotis californicus</i>	0.44%	G4G5T4	S3S4	CA
<i>Euphilotes pallescens arenamontana</i>	25.00%	G3?T1	S1	NV
<i>Gelochelidon nilotica</i>	31.82%	G5	S1	CA
<i>Gopherus agassizii</i>	0.11%	G3	S2S3	CA
<i>Gratiola heterosepala</i>	2.44%	G2	S2	CA
<i>Grusonia pulchella</i>	100.00%	G3G4	S3	NV
<i>Haliaeetus leucocephalus</i>	0.15%	G5	S3	CA
<i>Hesperia uncas fulvapalla</i>	100.00%	G4G5T1	S1	NV
<i>Horkelia cuneata var. puberula</i>	3.13%	G4T1	S1	CA
<i>Hulsea californica</i>	2.22%	G3	S3	CA
<i>Hulsea mexicana</i>	100.00%	G3	S1	CA
<i>Hydroprogne caspia</i>	33.33%	G5	S4	CA
<i>Icteria virens</i>	1.08%	G5	S3	CA
<i>Juga acutifilosa</i>	10.00%	G2?	S1	NV
<i>Larus californicus</i>	12.50%	G5	S4	CA
<i>Lasionycteris noctivagans</i>	1.41%	G3G4	S3	NV
<i>Lasiurus cinereus</i>	1.89%	G3G4	S2S3	NV
<i>Laterallus jamaicensis coturniculus</i>	0.51%	G3G4T1	S1	CA
<i>Legenere limosa</i>	3.23%	G2	S2	CA
<i>Lepidium integrifolium</i>	50.00%	G2G3T2T3	S1	NV
<i>Lepidurus packardi</i>	0.33%	G4	S3S4	CA
<i>Limnanthes alba ssp. parishii</i>	5.88%	G4T2	S2	CA
<i>Linanthus bellus</i>	2.13%	G2G3	S2	CA
<i>Linderiella occidentalis</i>	0.62%	G2G3	S2S3	CA
<i>Melospiza melodia</i>	0.07%	G5	S3?	CA
<i>Microdipodops pallidus</i>	8.57%	G3	S2	NV
<i>Myotis californicus</i>	2.00%	G5	S3S4	NV
<i>Myotis ciliolabrum</i>	4.81%	G5	S3S4	NV
<i>Myotis lucifugus</i>	3.13%	G3G4	S2S3	NV
<i>Myotis thysanodes</i>	1.96%	G4	S2	NV
<i>Myotis volans</i>	3.00%	G4G5	S3S4	NV
<i>Ochotona princeps</i>	1.27%	G5	S2	NV
<i>Oryctes nevadensis</i>	25.00%	G3	S2S3	NV
<i>Parastrellus hesperus</i>	4.10%	G5	S3S4	NV
<i>Pelecanus erythrorhynchos</i>	0.29%	G4	S1S2	CA
<i>Pelecanus occidentalis californicus</i>	16.67%	G4T3T4	S3	CA
<i>Penstemon albomarginatus</i>	4.55%	G2	S1	CA

<i>Petalonyx thurberi</i> ssp. <i>gilmanii</i>	25.00%	G5T2	S2	CA
<i>Plegadis chihi</i>	0.69%, 2.00%	G5	S3S4, S3B	CA, NV
<i>Polioptila melanura</i>	22.22%	G5	S3S4	CA
<i>Pyrgulopsis gibba</i>	2.78%	G3	S3	NV
<i>Pyrgulopsis lockensis</i>	100.00%	G1	S1	NV
<i>Pyrgulopsis ruinosa</i>	100.00%	GX	S1	NV
<i>Pyrgulopsis wongi</i>	25.00%	G2	S1	NV
<i>Rallus obsoletus yumanensis</i>	15.22%	G3T3	S1S2	CA
<i>Rana luteiventris</i> pop. 3	0.33%	G4T2T4Q	S2S3	NV
<i>Rynchops niger</i>	0.90%	G5	S2	CA
<i>Sagittaria sanfordii</i>	3.06%	G3	S3	CA
<i>Salvadora hexalepis virgultea</i>	3.57%	G5T4	S2S3	CA
<i>Selaginella eremophila</i>	1.61%	G4	S2S3	CA
<i>Sigmodon hispidus eremicus</i>	5.56%	G5T2T3	S2	CA
<i>Siphateles bicolor</i> ssp. 4	40.00%	G4T1Q	S1	NV
<i>Spea hammondii</i>	0.32%	G2G3	S3	CA
<i>Tetracoccus dioicus</i>	12.12%	G2G3	S2	CA
<i>Toxostoma lecontei</i>	1.82%	G4	S3	CA
<i>Vireo bellii pusillus</i>	0.24%	G5T2	S2	CA
<i>Vulpes macrotis</i>	100.00%	G4	S3	NV

Limestone monkeyflower (*Erythranthe calcicola*) in Nevada © Naomi Fraga





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